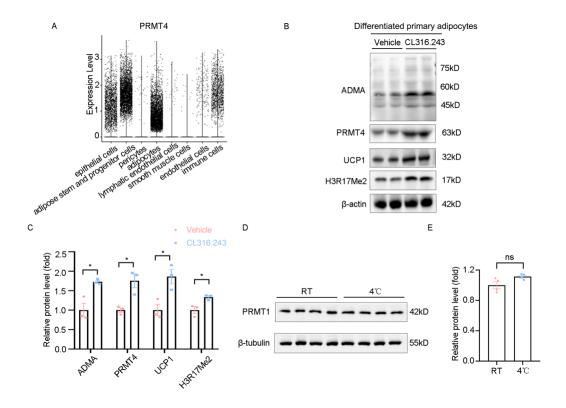
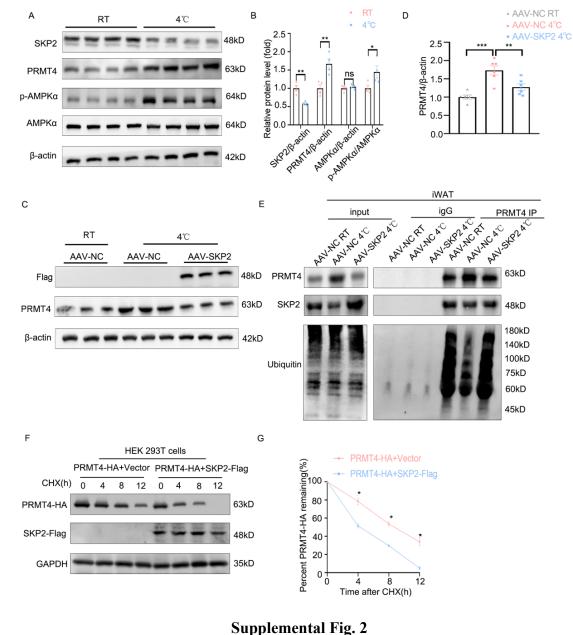
1	Supplemental Materials
2	PRMT4 facilitates white adipose tissue browning and thermogenesis by
3	methylating PPARγ
4	Short title: PRMT4 promotes WAT browning
5	Yi Zhong, Yilong Wang, Xiaoguang Li, Haojie Qin, Shu Yan, Caijun Rao, Di Fan,
6	Duqiu Liu, Fei Deng, Yanli Miao, Ling Yang, Kai Huang
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Supplemental Fig. 1

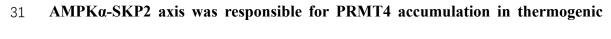
17 Levels of PRMT4 but not that of PRMT1 increased during iWAT browning.

A, Expression level of PRMT4 in each cell type of subcutaneous adipose tissue. B, SVF 18 cells were isolated from iWAT of C57BL/6J mice and differentiated for 6 days into 19 20 mature adipocytes, and then treated with CL316,243 (2 µM) for 4 h before harvested. Representative immunoblotting images of the expression levels of ADMA, PRMT4, 21 22 UCP1, and H3R17Me2 in the differentiated primary adipocytes. C, Quantitative analysis of the protein level of ADMA, PRMT4, UCP1, and H3R17Me2 in panel B. D, 23 Representative immunoblotting images displaying the expression levels of PRMT1 in 24 iWAT from mice hosted at RT or 4 °C for 24 h. E, Quantitative analysis of the protein 25 level of PRMT1 in panel D. Data are presented as mean \pm SEM. *P<0.05, ns, non-26 significant. 27



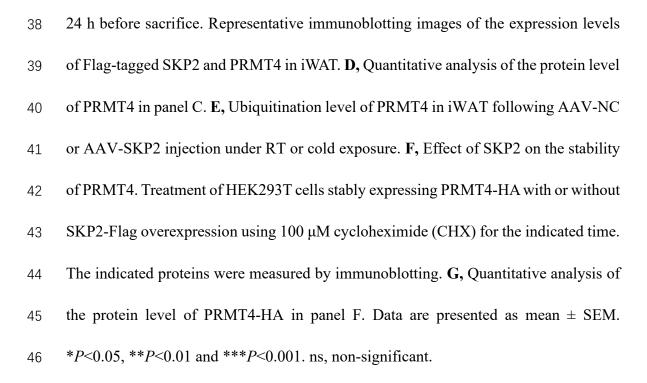


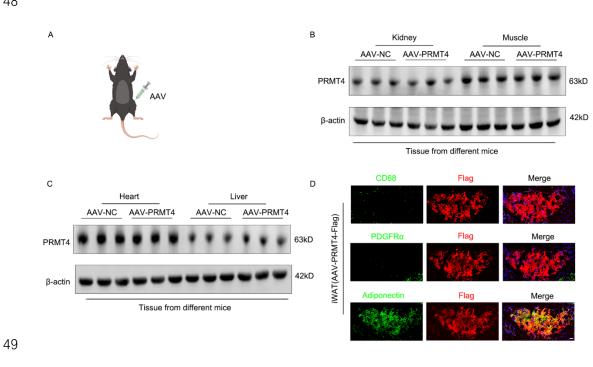
Suppremental Fig. 2



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32 program.
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A, Representative immunoblotting images of the expression levels of SKP2, PRMT4,
p-AMPKα, and AMPKα in iWAT from mice hosted at RT or 4 °C for 24 h. B,
Quantitative analysis of SKP2 and PRMT4 protein level as well as AMPKα
phosphorylation in panel A. C, AAV-NC or AAV-SKP2 were subcutaneously injected
vinto the bilateral inguinal areas, and mice were subjected to cold exposure (4°C) for



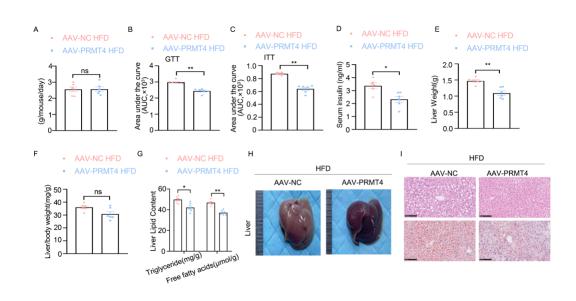


Supplemental Fig. 3

Specific overexpression of PRMT4 in iWAT.

A, Schematic diagram of local AAV injection subcutaneously into the bilateral inguinal areas. **B-C**, Representative immunoblotting images of the expression level of PRMT4 in the indicated tissues from C57BL/6J mice after injection of AAV-NC or AAV-PRMT4 in iWAT. β -actin protein was used as the loading control. **D**, Representative immunofluorescence images for Flag (red), CD68 (macrophage marker, green), PDGFR α (preadipocyte marker, green), and Adiponectin (adipocyte marker, green) in iWAT after injection of AAV-PRMT4-Flag. Nuclei staining by DAPI (blue). Scale bar, 20 µm.

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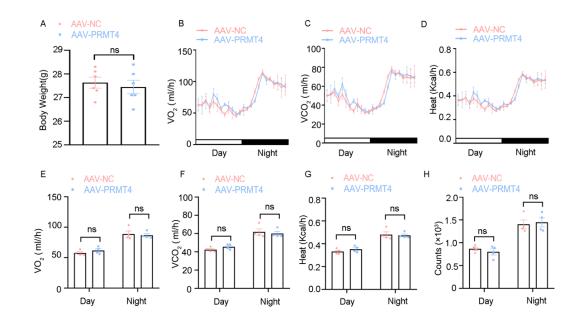
Supplemental Fig. 4

63 PRMT4 overexpression in iWAT meliorates diet-induced liver lipid accumulation. 64 AAV-NC or AAV-PRMT4 was subcutaneously injected into the bilateral inguinal areas, two weeks later, the mice received 8 weeks of HFD. A, Food intake of the mice. B, 65 Quantitative analysis of glucose tolerance test (GTT). C, Quantitative analysis of 66 insulin tolerance test (ITT). D, Fasting serum insulin levels after 8 weeks of HFD. E, 67 Liver weights of mice after 8 weeks of HFD. F, Liver/body weight ratio of mice after 68 69 8 weeks of HFD. G, Liver lipid content (triglycerides and free fatty acids) in mice. H, Gross morphology of the liver from mice after 8 weeks of HFD. I, Representative H&E 70

71 and Oil Red O staining of livers from mice after 8 weeks of HFD. Scale bar, 100 μm.

72 Data are presented as mean \pm SEM. **P*<0.05 and ***P*<0.01. ns, non-significant.

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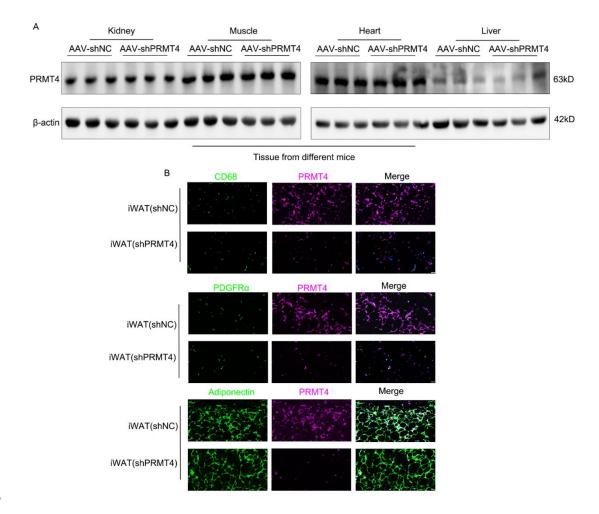
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Supplemental Fig. 5

76 **PRMT4 overexpression did not affect energy consumption at RT.**

AAV-NC or AAV-PRMT4 were subcutaneously injected via the bilateral inguinal areas. 77 78 A, Body weight of the mice was recorded. The whole-body oxygen consumption rate (VO₂) (ml/h) (**B**), carbon dioxide production (VCO₂) (ml/h) (**C**), and heat production 79 (D) of mice were measured using Columbus Oxymax metabolic chambers at RT. The 80 81 day/night bar represents a 12 h duration. E, Quantitative analysis of VO₂ in panel B. F, Quantitative analysis of VCO_2 in panel C. G, Quantitative analysis of heat production 82 in panel D. VO₂, VCO₂, and heat production were analyzed by ANCOVA with total 83 body mass as covariate. H, Average physical activity for 24 h. Data are presented as 84 mean \pm SEM. ns, non-significant. 85

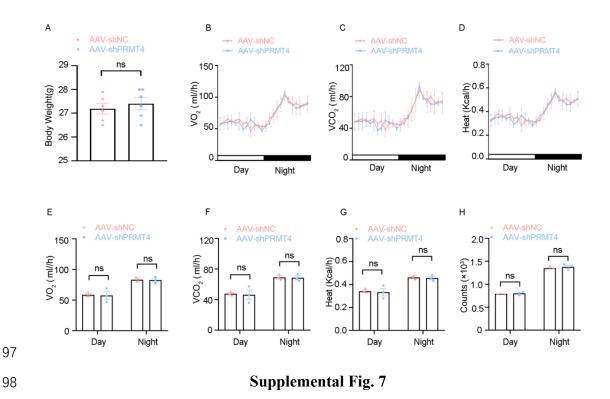


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Supplemental Fig. 6

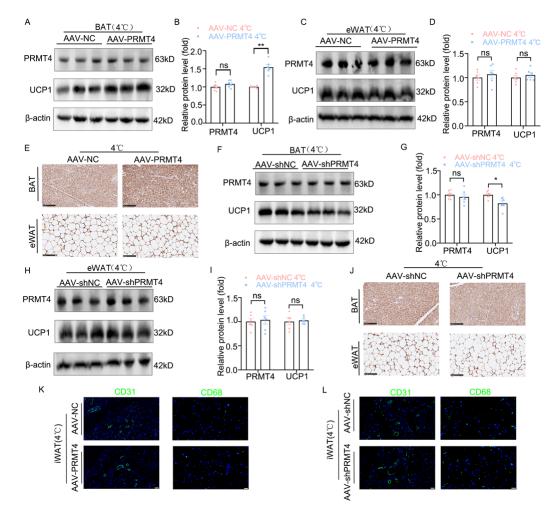
88 Specific knockdown of PRMT4 in iWAT.

89 **A**, Representative immunoblotting images of the expression level of PRMT4 in the 90 indicated tissues from C57BL/6J mice with iWAT injection of AAV-shNC or AAV-91 shPRMT4. β -actin protein was used as the loading control. **B**, Representative 92 immunofluorescence images for PRMT4 (purple), CD68 (macrophage marker, green), 93 PDGFR α (preadipocyte marker, green), and Adiponectin (adipocyte marker, green) in 94 iWAT after injection of AAV-shNC or AAV-shPRMT4. Nuclei staining by DAPI 95 (blue). Scale bar, 20 µm.

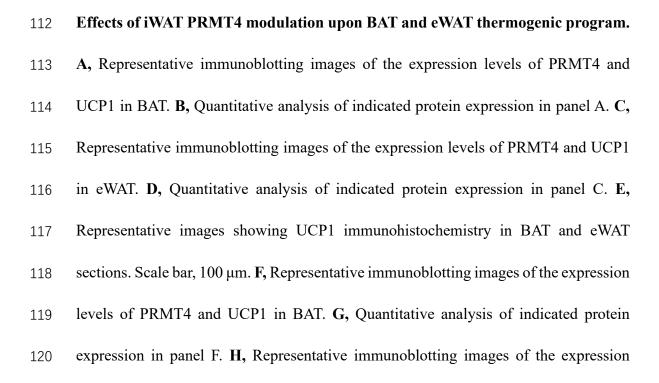




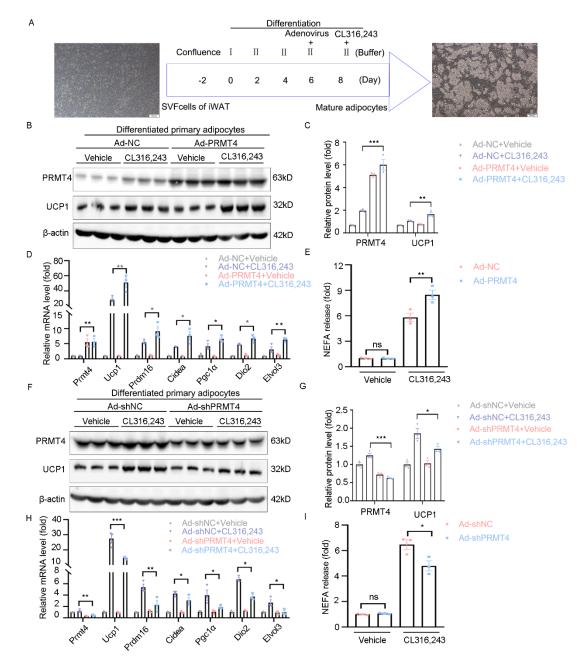
AAV-shNC or AAV-shPRMT4 were subcutaneously injected into the bilateral inguinal 100 101 areas. A, Body weight of the mice was recorded. The whole-body oxygen consumption rate (VO₂) (ml/h) (**B**), carbon dioxide production (VCO₂) (ml/h) (**C**), and heat 102 production (D) of mice were measured using Columbus Oxymax metabolic chambers 103 104 at RT. The day/night bar represents a 12 h duration. E, Quantitative analysis of VO₂ in panel B. F, Quantitative analysis of VCO₂ in panel C. G, Quantitative analysis of heat 105 production in panel D. VO₂, VCO₂, and heat production were analyzed by ANCOVA 106 with total body mass as covariate. H, Average physical activity for 24 h. Data are 107 presented as mean \pm SEM. ns, non-significant. 108



Supplemental Fig. 8



121	levels of PRMT4 and UCP1 in eWAT. I, Quantitative analysis of indicated protein
122	expression in panel H. J, Representative images showing UCP1 immunohistochemistry
123	in BAT and eWAT sections. Scale bar, 100 μ m. K, Representative image showing
124	immunofluorescence staining of CD31 (endothelial cell marker, green) or CD68
125	(macrophage marker, green) in iWAT with injection of AAV-NC or AAV-PRMT4
126	after cold exposure. Nuclei staining by DAPI (blue). Scale bar, 20 µm. L,
127	Representative image showing immunofluorescence staining of CD31 (endothelial cell
128	marker, green) or CD68 (macrophage marker, green) in iWAT with injection of AAV-
129	shNC or AAV-shPRMT4 after cold exposure. Nuclei staining by DAPI (blue). Scale
130	bar, 20 $\mu m.$ Data are presented as mean \pm SEM. *P<0.05 and **P<0.01. ns, non-
131	significant.



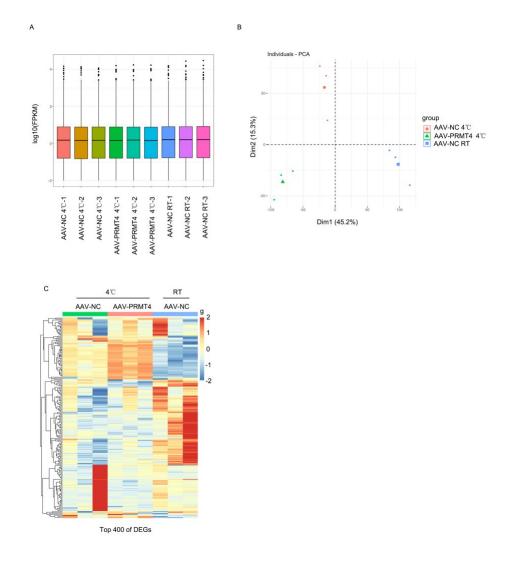
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Supplemental Fig. 9

135 **PRMT4 regulated thermogenic program of SVF cells.**

136 SVF cells were isolated from iWAT of C57BL/6J mice and differentiated for 6 days into 137 mature adipocytes, then the differentiated primary adipocytes were infected with the 138 indicated adenovirus for 48 h and treated with CL316,243 (2 μ M) for 4 h before 139 harvested. **A**, Schematic illustration of in vitro differentiation of SVF cells isolated from 140 iWAT. **B** and **F**, Representative immunoblotting images of the expression levels of

141	PRMT4 and UCP1 in the differentiated primary adipocytes after vehicle or CL316,243
142	administration. C, Quantitative analysis of the protein level of PRMT4 and UCP1 in
143	panel B. D and H, Relative mRNA levels of the indicated genes in the differentiated
144	primary adipocytes after vehicle or CL316,243 administration. E and I, Relative NEFA
145	levels in the culture medium. G, Quantitative analysis of the protein level of PRMT4
146	and UCP1 in panel F. Data are presented as mean ± SEM. *P<0.05, **P<0.01
147	and***P<0.001. ns, non-significant.

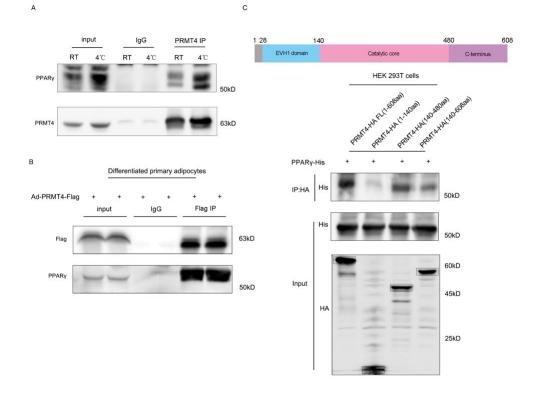


Supplemental Fig. 10

156 Identification of PRMT4-regulated genes by RNA-seq analysis.

157 **A**, The FPKM boxplot shows the distribution of gene expression levels among each

- 158 sample. B, Principal component analysis (PCA) revealed global gene expression
- 159 patterns upon cold stimulation or PRMT4 transduction in iWAT. C Hierarchical
- 160 clustering revealed the top 400 differentially expressed genes (DEGs).

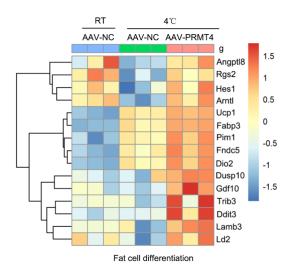


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Supplemental Fig. 11

163 **PRMT4 interacted with PPARγ.**

164 **A,** iWAT tissues from mice were hosted at RT or 4 °C for 24 h, co-IP assay was 165 conducted. **B,** SVF cells were isolated from iWAT of C57BL/6J mice and differentiated 166 for 6 days into mature adipocytes, then the differentiated primary adipocytes were 167 infected with Ad-PRMT4-Flag for 48 h and followed by co-IP assay. **C,** The interaction 168 domains of PRMT4 with PPAR γ were explored using full-length and deletion-mutated 169 PRMT4 expression constructs based on co-IP assays.

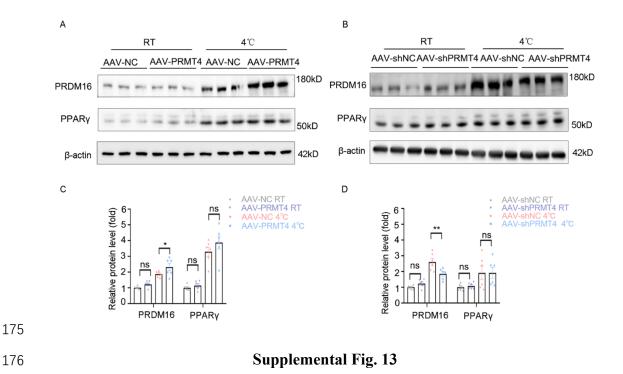


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Supplemental Fig. 12

172 **DEGs were enriched in fat cell differentiation**

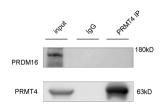
173 Fat cell differentiation-related DEGs were shown. Heatmaps represents gene174 expression changes.



177 **PRMT4 regulated cold-induced PRDM16 expression in iWAT.**

178 A, B, Representative immunoblotting images showing the expression levels of

179 PRDM16 and PPAR γ in iWAT from mice injected with the indicated virus 180 subcutaneously into bilateral inguinal areas. **C-D**, Quantitative analysis of the protein 181 level of PRDM16 and PPAR γ in panel A and B. Data are presented as mean \pm SEM. 182 ***P*<0.01 and****P*<0.001, ns, non-significant. 183



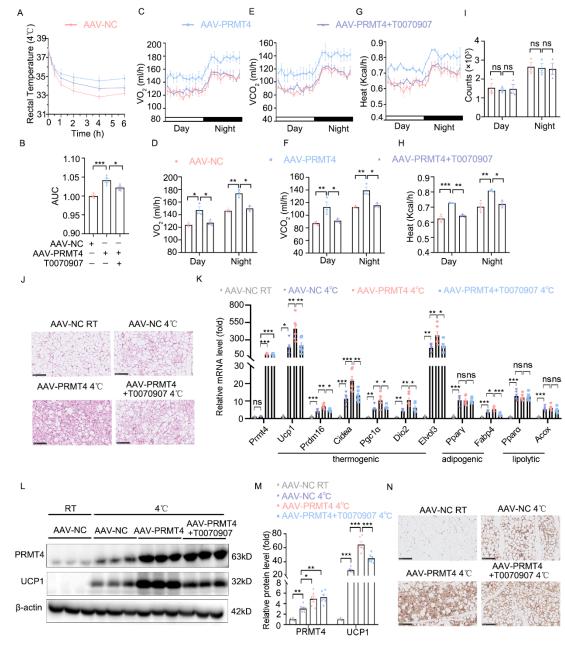
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Supplemental Fig. 14

186 **PRMT4 did not directly interact with PRDM16**

187 Co-IP assay showed that PRMT4 did not bound to PRDM16 in iWAT.





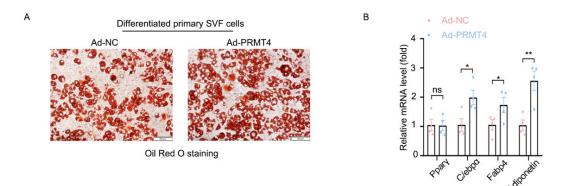
Supplemental Fig. 15

191 Inhibition of PPARγ by T0070907 abolishes the effect of PRMT4 on thermogenic

192 gene expression.

193 **A**, The rectal temperature of mice during acute cold exposure (4°C) was monitored in 194 the first 6 h. **B**, Quantitative analysis of rectal temperature in panel A. The oxygen 195 consumption rate (VO₂) (**C**), carbon dioxide production (VCO₂) (**E**), and heat 196 production (**G**) were analyzed. The day/night bar represents a 12 h duration. **D**,

197	Quantitative analysis of VO_2 in panel C. F, Quantitative analysis of VCO_2 in panel E.
198	H, Quantitative analysis of heat production in panel G. VO ₂ , VCO ₂ , and heat production
199	were analyzed by ANCOVA with total body mass as covariate. I, Average values of
200	physical activity. J, Representative HE staining of iWAT sections; scale bar, 100 μ m.
201	K, Relative mRNA levels of thermogenic, adipogenic, and lipolytic genes. L,
202	Representative immunoblotting images of the expression levels of PRMT4 and UCP1
203	in iWAT. M, Quantitative analysis of indicated protein level in panel L. N,
204	Representative IHC images of UCP1 in iWAT sections were shown. Scale bar, 100 μ m.
205	Data are presented as the mean \pm SEM. * <i>P</i> <0.05, ** <i>P</i> <0.01 and *** <i>P</i> <0.001. ns, non-
206	significant.



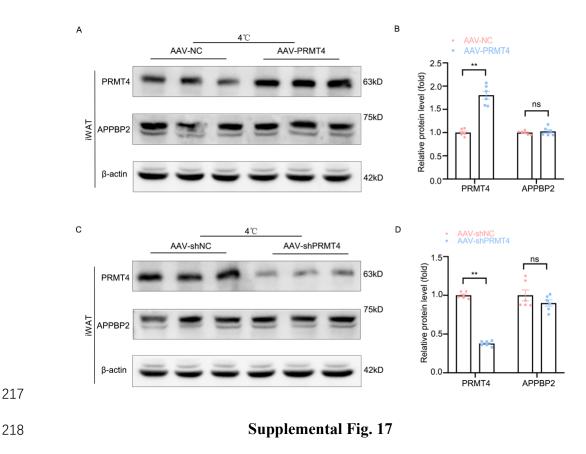
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Supplemental Fig. 16

210 **PRMT4 overexpression promotes adipogenesis in vitro.**

SVF cells isolated from iWAT of C57BL/6J mice were infected with the indicated adenovirus and were then differentiated for 6 days into mature adipocytes in vitro. **A**, Representative Oil-red O staining of differentiated adipocytes on day 6. Scale bar, 100 μ m. **B**, Relative mRNA levels of the indicated genes in differentiated adipocytes on day 6. Data are presented as mean \pm SEM. **P*<0.05, ***P*<0.01. ns, non-significant.



219 PRMT4 overexpression or knockdown did not affect protein level of APPBP2 in
220 iWAT upon cold exposure.

AAV-NC, AAV-PRMT4, AAV-shNC or AAV-shPRMT4 was subcutaneously injected into the bilateral inguinal areas, two weeks later, mice were subjected to cold exposure (4°C) for 24 h before sacrifice. **A&C**, Representative immunoblotting images of the expression levels of PRMT4 and APPBP2 in iWAT. **B&D**, Quantitative analysis of indicated protein expression in panel A&C. Data are presented as mean \pm SEM. ***P*<0.01, ns, non-significant.

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233 Table S1. Primers for qPCR

Gene	Forward 5' \rightarrow 3'	Reverse 5'→3'
h-β-actin	GGCACCCAGCACAATGAA	GGAAGGTGGACAGCGAGG
h-Prmt4	TCGCCACACCCAACGATTT	GTACTGCACGGCAGAAGACT
h-Ucp1	AGGTCCAAGGTGAATGCCC	TTACCACAGCGGTGATTGTTC
m-18S	TTGACGGAAGGGCACCACCAG	GCACCACCACCACGGAATCG
m-Prmt4	TGACATCAGTATTGTGGCACAG	CTGAGGAGCCTAAGGGAATCA
m-Ucp1	AGGCTTCCAGTACCATTAGGT	CTGAGTGAGGCAAAGCTGATTT
m-Pgc1a	GGTTGAAAAAGCTTGACTGGCG	ACCAACCAGAGCAGCACACT
m-Elvol3	TTCTCACGCGGGTTAAAAATGG	TCTCGAAGTCATAGGGTTGCAT
m-Prdm16	CCAAGGCAAGGGCGAAGAA	AGTCTGGTGGGATTGGAATGT
m-Dio2	AATTATGCCTCGGAGAAGACCG	GGCAGTTGCCTAGTGAAAGGT
m-Pparα	AGAGCCCCATCTGTCCTCTC	ACTGGTAGTCTGCAAAACCAAA
m-Acox	AAATATGCCCAGGTGAAGCC	CACTGTATCGAATGGCAATGG
m-Ppary	TCGCTGATGCACTGCCTATG	GAGAGGTCCACAGAGCTGATT
m-Fabp4	AAGGTGAAGAGCATCATAACCCT	TCACGCCTTTCATAACACATTCC
m-Adiponectin	TGTTCCTCTTAATCCTGCCCA	CCAACCTGCACAAGTTCCCTT
m-C/ebpα	CAAGAACAGCAACGAGTACCG	GTCACTGGTCAACTCCAGCAC