Supplementary Figures

Leptin signaling suppression in macrophages improves immunometabolic outcomes in obesity

Lauar de Brito Monteiro¹, Juliana Silveira Prodonoff¹, Cristhiane Favero de Aguiar¹, Felipe Correa-da-Silva¹, Angela Castoldi², Nikki van Teijlingen Bakker³, Gustavo Gastão Davanzo¹, Bianca Castelucci¹, Jéssica Aparecida da Silva Pereira^{1,8}, Jonathan Curtis^{3, 4}, Jörg Büscher³, Larissa Menezes dos Reis¹, Gisele Castro¹, Guilherme Ribeiro¹, João Victor Virgílio-da-Silva¹, Douglas Adamoski⁵, Sandra Martha Gomes Dias⁵, Silvio Roberto Consonni⁶, Jose Donato Jr⁷, Edward J. Pearce^{3,4}, Niels Olsen Saraiva Câmara⁸, Pedro M. Moraes-Vieira^{1, 9, 10*}

¹Laboratory of Immunometabolism, Department of Genetics, Evolution, Microbiology and Immunology, University of Campinas, Brazil.

² Laboratory Keizo Asami, Immunopathology Laboratory, Federal University of Pernambuco, Pernambuco, Brazil.

³ Department of Immunometabolism, Max Planck Institute of Epigenetics and Immunobiology, Freiburg im Breisgau, Germany.

⁴ Bloomberg Kimmel Institute, and Department of Oncology, Johns Hopkins University School of Medicine, Baltimore, USA.

⁵ Brazilian Biosciences National Laboratory (LNBio), Brazilian Center for Research in Energy and Materials (CNPEM), Campinas, Sao Paulo, Brazil.

⁶ Department of Biochemistry & Tissue Biology, Institute of Biology, University of Campinas, Campinas, Brazil.

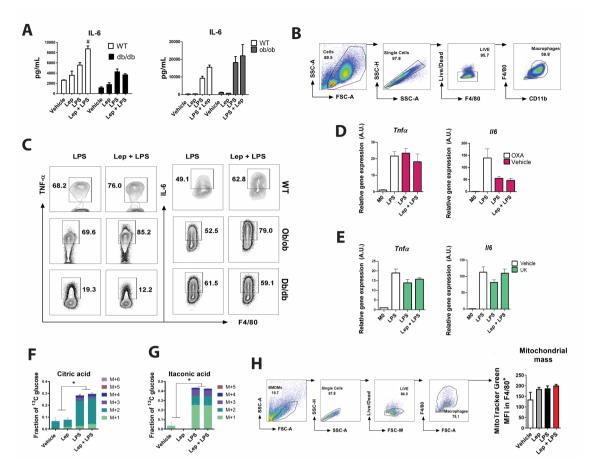
⁷ Department of Physiology and Biophysics, Institute of Biomedical Sciences, University of São Paulo, São Paulo, Brazil.

⁸ Department of Immunology, Institute of Biomedical Sciences IV, University of São Paulo, São Paulo, Brazil.

⁹ Experimental Medicine Research Cluster (EMRC), University of Campinas, São Paulo, Brazil.

¹⁰ Obesity and Comorbidities Research Center (OCRC), University of Campinas, São Paulo, Brazil.

^{*} Corresponding author: Prof. Dr. Pedro M M Moraes-Vieira, Department of Genetics, Evolution, Microbiology and Immunology, Institute of Biology, University of Campinas, São Paulo, Brazil. E-mail: <u>pmvieira@unicamp.br</u>



Supplementary Figure 1. Leptin directly modulates macrophage metabolism.

A) Concentration of IL-6 in the supernatant of leptin/LPS activated BMDMs from wild type (WT), db/db, or ob/ob mice by ELISA;

B) Flow cytometry gating Strategy for analysis of live peritoneal macrophages;

C) TNF- α and IL-6 positive macrophages from peritoneal lavage of WT, ob/ob and db/db mice were analyzed by flow cytometry;

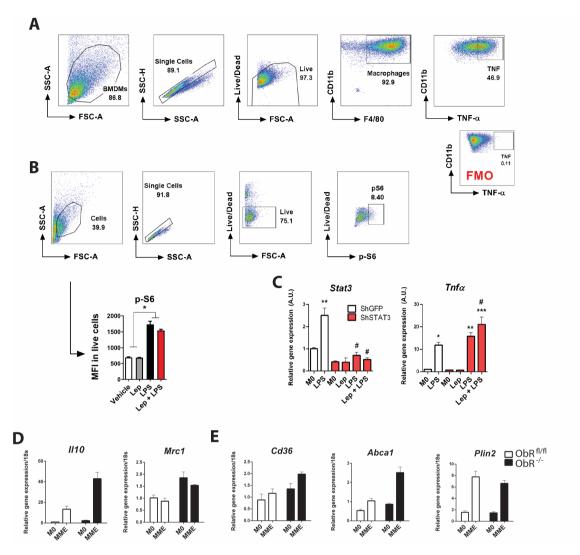
D) Relative gene expression of $Tnf\alpha$ and *II6* from BMDMs treated with 40 mM oxamate (OXA), and/or leptin+LPS for 6 hours.

E) Relative gene expression of *Tnf* α and *II6* from BMDMs treated with 10µM UK-5099 (UK), and/or leptin+LPS for 6 hours.

F-G) Fraction contribution of glucose carbons into citrate and G) itaconate, in macrophages stimulated with leptin/LPS for 6 hours and cultured in ¹³C-glucose containing medium;

H) Flow cytometry gating strategy and Median of Fluorescence Intensity (MFI) of MitoTracker Green positive BMDMs after 6 hours of activation with leptin/LPS;

Data are represented as mean values \pm s.e.m. n=3/5 per group. One-way ANOVA with Bonferroni's multiple comparison test (A, D-H) *p<0.05; #p<0.05 compared to all groups; Data are representative of three/two independent experiments.



Supplementary Figure 2. Strategies for studying leptin signaling in macrophages.

A) Representative flow cytometry gating strategy of cytokines (in this case, TNF- α) positive BMDMs after 6 hours of activation with leptin/LPS (100 ng/mL);

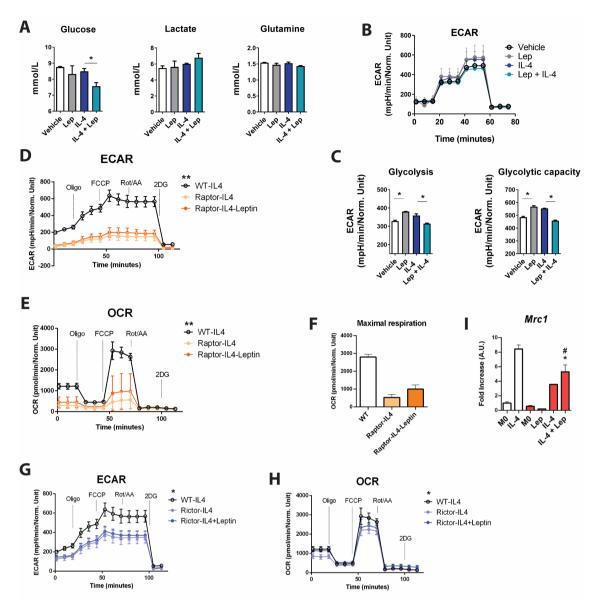
B) Representative flow cytometry gating strategy and MFI of phospho-S6 positive BMDMs after 30 min activation with leptin/LPS;

C) Relative gene expression of *Stat3* and *Tnf* α by shSTAT3 or shGFP (control) transduced BMDMs;

D) Relative gene expression of II10 and Mrc1 by metabolically activated macrophages (MMe);

E) Relative gene expression of Cd36, Abca1 and Plin2 in metabolically activated macrophages (MMe);

Data are represented as mean values \pm s.e.m. (C, F). n=3/5 per group. One-way ANOVA with Bonferroni's multiple comparison test (C, E). A,B representative of three independent experiments, C-F representative of two independent experiments. MFI: Median of Fluorescence Intensity.



Supplementary Figure 3. Metabolic effects of leptin on M(IL-4).

A) Concentration of Glucose, Lactate and Glutamine concentrations in the supernatant of BMDMs were measured by Bioanalyser;

B) Extracellular acidification rate (ECAR) of macrophages upon glycolytic stress (injections of glucose, oligomycin and 2-DG);

C) ECAR analysis of glycolysis (followed by glucose injection); Glycolytic capacity (followed by oligomycin injection).

D) Extracellular acidification rate (ECAR) of IL-4-treated macrophages from WT and Raptor⁻ mice upon glycolytic stress (injections of glucose, oligomycin and 2-DG);

E) Oxygen consumption rate (OCR) of IL-4-treated macrophages from WT and Raptor^{-/-} mice upon mitochondrial stress test (injections of oligomycin, FCCP, antimycin and rotenone);

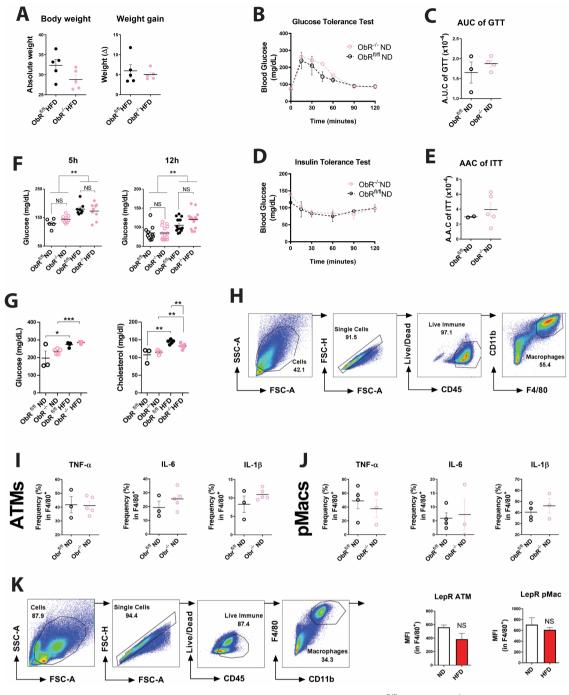
F) Maximal respiration (followed by FCCP injection) of IL-4-treated macrophages from WT and Raptor^{-/-} mice;

G) Extracellular acidification rate (ECAR) of IL-4-treated macrophages from WT and Rictor^{-/-} mice upon glycolytic stress (injections of glucose, oligomycin and 2-DG);

H) Oxygen consumption rate (OCR) of IL-4-treated macrophages from WT and Rictor^{-/-} mice upon mitochondrial stress test (injections of oligomycin, FCCP, antimycin and rotenone);

I) Relative gene expression of Mrc1 by shSTAT3 or shGFP (control) transduced BMDMs.

Data are represented as mean values \pm s.e.m. n=3/5 per group. One-way ANOVA with Bonferroni's multiple comparison test. A-H representative of two to three independent experiments.



Supplementary Figure 4. Metabolic and immune evaluation of ObR^{fl/fl} and ObR^{-/-} mice fed with chow and high fat diet.

A) Absolute body weight (g) and weight gain (Δ) of ObR^{fl/fl} mice and ObR^{-/-} mice fed a high fat diet (HFD) over the course of 16 weeks;

- B) Insulin Tolerance Test (ITT) in chow-fed ObR^{fl/fl} and ObR^{-/-} mice;
- C) Area under the curve (AUC) of ITT test in chow-fed ObR^{fl/fl} and ObR^{-/-} mice;
- D) Insulin Tolerance Test (ITT) in chow-fed ObR^{fl/fl} and ObR^{-/-} mice;
- E) Area above the curve (AAC) of ITT test in chow-fed ObR^{fl/fl} and ObR^{-/-} mice
- F) Glucose levels in the blood after 5 and 12 hours of fasting;
- G) Glucose and cholesterol levels in the serum of ND- and HFD-fed ObR^{fl/fl} and ObR^{-/-} mice;

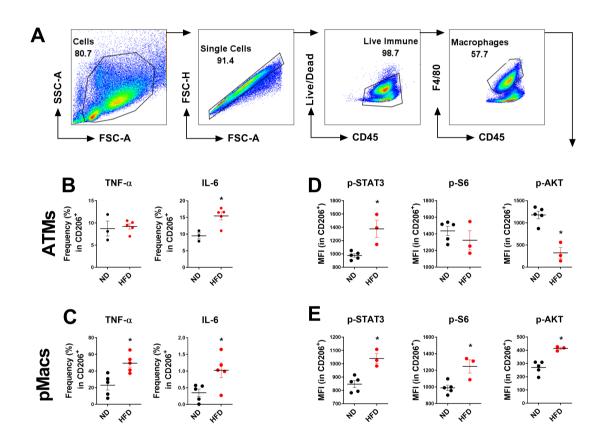
H) Flow cytometry gating strategy for evaluation of live macrophages;

I) Frequency of IL-6, IL-1 β and TNF- α positive macrophages (F4/80⁺) from the AT of chowfed ObR^{fl/fl} and ObR^{-/-} mice;

J) Frequency of IL-6, IL-1 β and TNF- α positive macrophages (F4/80⁺) from the peritoneal lavage of chow-fed ObR^{fl/fl} and ObR^{-/-} mice;

K) Flow cytometry gating strategy for macrophage isolation and MFI analysis of ObR expression in ATMs and pMacs of chow- (normal diet - ND) and HFD-fed ObR reporter mice.

Data are represented as mean values ± s.e.m. n=3/5 per group. Two-way ANOVA Sidak's multiple comparison test Unpaired (b, d). Two-tailed Student's t test (A, C, E, I-K). One-way ANOVA with Bonferroni's multiple comparison test (F-G). A-K representative of two to three experiments. MFI: Median of Fluorescence Intensity.



Supplementary Figure 5. High fat diet impact on AT and peritoneal CD206⁺ macrophages.

A) Flow cytometry representative gating strategy to select macrophages from the AT and peritoneal lavage;

B) Frequency of TNF- α and IL-6 positive CD206⁺ macrophages from the AT of mice fed a chow normal diet (ND) or a high fat diet (HFD);

C) Frequency of TNF- α and IL-6 positive CD206⁺ macrophages from the peritoneal lavage of mice fed a chow normal diet (ND) or a high fat diet (HFD);

D) MFI of phospho-STAT3, phospho-S6 and phospho- AKT^{ser473} in CD206⁺ macrophages from the AT of mice fed a chow normal diet (ND) or a high fat diet (HFD);

E) MFI of phospho-STAT3, phospho-S6 and phospho- AKT^{ser473} in CD206⁺ macrophages from the peritoneal lavage of mice fed a chow normal diet (ND) or a high fat diet (HFD).

Data are represented as mean values \pm s.e.m. n=3/5 per group. Unpaired two-tailed Student's t test (B-E), *p<0.05. Data are representative of three experiments. MFI: Median of Fluorescence Intensity.