

## **Supplementary Appendix**

Supplement to: **Vibha Anand et al., Islet Autoimmunity and HLA Markers of Presymptomatic and Clinical Type 1 Diabetes: Joint Analyses of Prospective Cohort Studies in Finland, Germany, Sweden, and The United States**

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## **Study Sites, Newborn Screening and Recruitment**

**The BABYDIAB/BABYDIET study:** is a prospective, longitudinal birth-cohort study in Germany (1,2). Between 1989 and 2000, BABYDIAB enrolled 1,650 offspring of mothers and/or fathers with type 1 diabetes. Between 2000 and 2006, BABYDIET enrolled 791 offspring or siblings of patients with type 1 diabetes. Participants were enrolled at birth and followed with type 1 diabetes-related autoantibody testing at age 9 months, at 2 years, and every 3 years thereafter. Those who became islet autoantibody-positive were subsequently followed six-monthly. A subgroup of 150 children participated in the BABYDIET dietary intervention study and had 3-monthly follow-up visits until the age of 3 years, and yearly thereafter (ClinicalTrials.gov Identifier: NCT01115621). The intervention (delay of exposure to gluten) failed to show an effect on islet autoimmunity or type 1 diabetes risk. After completion of the trial, participants were followed further in BABYDIAB/BABYDIET. Written informed consent was provided by participating families. BABYDIAB was approved by the Bavarian ethical committee (Bayerische Landesärztekammer; No. 95357) and the BABYDIET was approved by the ethics committee of the Ludwig-Maximilians-University, Munich, Germany (No. 329/00).

**Diabetes Autoimmunity Study in the Young (DAISY):** is a prospective, longitudinal study located in Denver, Colorado. Screening for genetic risk was conducted using cord blood collected from 32,114 general population infants born at Denver's St. Joseph hospital between 1994 and 2004. Excluded were extremely premature newborns and those with severe congenital malformation or disease. Upon completion of informed consent, cord blood samples were typed for type 1 diabetes -associated HLA-DR, DQ genotypes. (3) Children with high-risk, (DRB1\*04-DQB1\*03:02/DRB1\*03) or moderate-risk (DRB1\*04-DQB1\*03:02/ DRB1\*04-DQB1\*03:02, DRB1\*04-DQB1\*03:02/DRX, DRB1\*03/DRB1\*03) or a specific low-risk (DRB1\*04-DQB1\*0302/DR2) genotypes were invited to participate in the follow-up protocol. (4) In 1993-95, first-degree relatives of individuals with type 1 diabetes were also enrolled, regardless of HLA genotype, if younger than 8 years. Starting in 1996, the eligibility for relatives was restricted to those younger than 4 years. Participants were enrolled into the follow-up study if informed consent was completed and at least one clinic visit, with type 1 diabetes-related autoantibody testing, was completed. All children enrolled at birth were tested at 9, 15, and 24 months of age and annually thereafter for islet autoantibodies. Children who were autoantibody positive were placed on an accelerated schedule in which they returned for a blood draw every 3 to 6 months.

**Diabetes Evaluation in Washington Study (DEWIT):** The DEW-IT study is a prospective, population-based observational study that HLA-screened 42,000 Washington State newborns born from 1995-2001 and 2008-2012 (5), under IRB approval and informed consent, using dried bloodspots from the Washington State Department of Health Newborn Screening Program. HLA-DQ genotyping utilized direct sequencing of exon 2 of each of DQA1 and DQB1. Individuals were eligible for further study if they had at least one of two high risk HLA haplotypes (DQA1\*03-DQB1\*03:02 and/or DQA1\*05:01-DQB1\*02:01) plus no protective haplotype (DQA1\*01-DQB1\*05:03, DQA1\*01-DQB1\*06:01, DQA1\*01-DQB1\*06:02, DQA1\*01-DQB1\*06:03, DQA1\*05-

DQB1\*0301, DQA1\*03-DQB1\*03:01, DQA1\*02-DQB1\*03:03 or DQA1\*02-DQB1\*02:02). Based on HLA genotyping, 14.2% of the screened population was deemed at increased risk for developing type 1 diabetes (estimated to contain 68% of future cases). Eligible children were offered periodic surveillance for three islet autoantibodies (GADA, IA2A and IAA) by mail-based or provider-based sampling, via testing in a DASP/IASP participating laboratory under CLIA certification using a DASP/IASP participating laboratory under CLIA certification using published methods (6). A total of 4,674 individuals were tested, with >10% having at least one islet autoantibody present. Children positive for any autoantibodies were asked to submit additional sample(s) for confirmation and closer surveillance. Children with no autoantibodies were followed less frequently. Web-based methods for contact, conveying antibody test results and education about early disease symptoms were also developed. The primary objective of DEW-IT has been to develop methods for population-based type 1 diabetes prediction that are accurate, cost-efficient and suitable to incorporate into the existing healthcare infrastructure. However, because all contact was remote (by phone and mail) the follow up sampling was often less frequent than in the other studies.

**Diabetes Prediction in Skane (DiPiS):** The DiPiS study is a prospective, longitudinal, population-based study in the southern part of Sweden. (7) Cord blood samples were obtained from 35,688 children born between September 2000 and August 2004 and analysed for type 1 diabetes HLA risk genotypes and cord-blood islet autoantibodies. HLA DQ genotypes were estimated as high-risk (DQA1\*05-B1\*02/DQB1\*03:02 (6 points)), medium-risk (DQB1\*03:02/DQB1\*06:04 or DQB1\*03:02/X (4 points)), low-risk (DQA1\*05-B1\*02/DQB1\*03:04, DQA1\*05-B1\*02/DQB1\*06:04, DQA1\*05-B1\*02/X (3 points)), neutral (2 points) or protective (0 points).(8) Based on prior observations other risk factors were included in the final risk score: first-degree relative (2 points), positive for cord blood islet autoantibodies (2 points), ABO incompatibility (1 point), febrile infection during pregnancy (1 point), presence of severe life event (1 point), high or low maternal age (1 point) as well as born large for gestational age (1 point). Parents were asked to participate if their child reached 3 points or more. A total of 7,826 families were invited to participate and follow-up samples from 4359 children have been included in the 15-year follow-up. Among the participating children, 233 (12.4 %) had a high-risk HLA genotype, 562 (30 %) medium-risk, 488 (26 %) low-risk, 524 (30 %) neutral, and 67 (3.6 %) had protective genotype. Initial follow up was performed yearly with islet autoantibodies and a yearly questionnaire regarding lifestyle, medical, psychological and dietary factors. Children who developed islet autoimmunity with two or more islet autoantibodies transferred to an intensive follow-up group and were followed every three months with islet autoantibodies, random blood glucose, HbA1c, height and weight and were offered a yearly oral glucose tolerance test.(9)

**Diabetes Prediction and Prevention Study (DIPP):** The DIPP study is a prospective, population-based birth cohort study launched in 1994 in Finland, where it is ongoing in Oulu, Tampere and Turku University Hospitals (10). In 1994-2009 a total of 145607 children were screened for HLA-DR/DQ associated susceptibility to type 1 diabetes from cord blood using sequence-specific oligonucleotide probes as previously described (11,12). The majority of the children eligible for follow-up carried either the high-risk HLA

genotype DQB1\*02/\*0302 or the moderate-risk genotypes DQB1\*0302/x (x≠\*02, \*0301, \*0602, or \*0603). In addition, a few other HLA genotypes were included as detailed in the Supplementary Table 1 of Ilonen et al. (13) A total of 17572 children were eligible and 13204 were enrolled in the study after signing the informed consent. Children who had at least three follow-up visits (n= 11680) were included in the present study.

DIPP participants had follow-up visits at the Oulu and Tampere centers at 3, 6, 12, 18 and 24 months of age and annually thereafter while in the Turku center the visits occurred every 3 months until 2 years of age and biannually thereafter. Children with any positive islet autoantibody were followed 3-monthly until 15 years of age or the diagnosis of type 1 diabetes. Height and weight were measured, and venous blood sample was drawn at every visit. In addition, clinical data such as diet, diseases, medications and vaccinations were collected by using diaries and interviews during study visits. (14,15)

## **Measurement of islet autoantibodies**

### **BABYDIAB/BABYDIET:**

IAA, GADA, IA-2A, and ZnT8A were measured in venous blood samples by the Institute of Diabetes Research, Helmholtz Center Munich, using radiobinding assays (RBAs) and thresholds based on the upper 99th centile and Q-Q plots of results from control children as described elsewhere (1,16,17). IAA levels were measured using a competitive RBA. (18) Assays (laboratory 121) were evaluated by the Diabetes Antibody Standardization Program (DASP) and the Islet Autoantibody Standardization Program (IASP). (19–22)

### **DAISY:**

All antibodies were measured, in duplicate, from frozen serum samples. Autoantibody detection assays have modified over the course of the 24 years study history. Methodology and time periods are described below for each autoantibody measured by the DAISY Study.

### **Glutamic acid decarboxylase antibodies (GADA) and Islet antigen-2 antibodies (IA-2A):**

June 8, 1994-Feb. 16, 2010 autoantibodies to GAD65 were measured utilizing a modified radioassay with in vitro transcribed and translated human GAD65 labeled with [<sup>3</sup>H]leucine. (23,24) (Results expressed as an index: (sample cpm-negative control cpm)/(positive control cpm-negative control cpm). Samples which test positive for autoantibodies are re-tested for confirmation, using a separately aliquoted sample from the same clinic visit blood draw.

Nov. 17, 1995-Feb. 16, 2010 autoantibodies to islet cell antigen 512 (IA-2) were measured utilizing a modified radioassay with in vitro transcribed and translated human ICA512bdc protein construct labeled with [<sup>35</sup>S]methionine. (24,25) The GADA and IA-2A autoantibodies were measured in a combined assay. Samples which test positive for autoantibodies are re-tested for confirmation, using a separately aliquoted sample from the same clinic visit blood draw.

Feb. 22, 2010-present the GADA and IA-2A assays were standardized to be brought into alignment with Bristol, UK and Munich, Germany labs. (17)

**Insulin autoantibodies (IAA):**

March 15, 1994-Dec. 18, 1998 insulin autoantibodies (IAA) were measured using a quantitative radioimmunoassay, competitive insulin assay (cIAA), utilizing [<sup>125</sup>I] labeled and unlabeled insulin (26) and precipitation with polyethylene glycol. This assay requires a substantial amount of serum, 600ul, so testing was not routinely repeated.

Jan. 27, 1999-present IAA is measured using a micro-IAA assay (mIAA), which utilizes ELISA techniques and Protein A-Sepharose with a much decreased volume of serum and reagents. (27) The assay is conducted in a 96-well format to allow for high-throughput analysis and may be repeated for confirmation of positive samples.

**Zinc transporter 8 autoantibodies (ZnT8A)**

Dec. 2010-present ZnT8 autoantibodies are measured for islet antibody positive participants. Detection of ZnT8 autoantibodies utilizes the radioimmunoassay similar to GADA and IA-2A, with modifications to allow for the ZnT8 probe. The ZnT8 probe is a synthetic molecule fusion cDNA that is transcribed in vitro and translated in the presence of [<sup>35</sup>S]methionine. (28,29)

ZnT8 was measured in all ever islet antibody positive participants. If the sample was positive for ZnT8, frozen serum samples from previous clinic visits were tested for ZnT8. All islet antibody positive participants were also tested for ZnT8 antibodies going forward. In 2012, all participants who completed a blood draw at a clinic visit, were tested for ZnT8 antibodies.

**DEWIT:**

Whole Glutamate Decarboxylase (GAD65) or the intracellular portion of insulinoma antigen-2 (IA2) were biosynthetically radiolabeled in the presence of <sup>35</sup>S-methionine, while HPLC-purified <sup>125</sup>I-monoiodo-Tyr<sub>A14</sub>-human insulin was purchased commercially (Perkin-Elmer, Hopkinton MA USA). Each antigen was incubated separately in triplicate wells with 2 to 5 uL of patient serum, then captured by Protein A, washed extensively to remove unbound antigen, and counted. Results were expressed using an index relative to specific positive control and negative control sera as described, with each cutoff defined as the 99<sup>th</sup> percentile of >200 individual healthy population controls. (6) When measured, ZnT8 antibodies were detected via an analogous procedure using biosynthetically <sup>35</sup>S-methionine radiolabeled antigen expressed from a construct comprising a hinged dimer containing the antigenic peptide loop of each of the aa325-Arg and the aa325-Trp alleles (JH6.2). Samples close to the cutoff, and those where triplicate well CV's were >30%, were reflexively repeated. The laboratory participates in a DASP/IASP proficiency testing, with DEW-IT assays exceeding mean performance of all participating laboratories for all 4 antigens over each of 12 consecutive workshops. The four autoantibodies assays are each CLIA certified (CLIA# 50D0982418).

**DiPiS:****Autoantibodies to GAD65 and IA-2: Analysis in dried blood spots, serum or plasma**

GAD65 autoantibodies (GADA) and IA-2 autoantibodies (IA2A) were analyzed in dried blood spots (DBS) with a radioligand binding assay (RBA). (49,50) GADA and IA2A levels were expressed as units per mL (U/mL) derived from the WHO standard 97/550.

The samples were considered to be positive if the IA2A levels were above 5 U/mL or the GADA levels were above 34 U/mL.

#### **Autoantibodies to IAA**

Analysis for IAA was first performed using a Noncompetitive method where serum samples (7 mL) were added to duplicate microplate wells and 125I was added and incubated on a shaker at 4°C for 48 h. PAS in a 40% slurry was added to a filter plate and washed three times with Tris buffer. Supermix scintillation solution was added to the wells after the plate had dried for 15 min. The radioactivity was measured in a  $\beta$ counter (Wallac Micro Beta Trilux; PerkinElmer). Positive samples for IAA were further analyzed using a competitive method. Serum samples were added to four wells in a 96-well plate. To these wells, 36 mL of 125I insulin with an activity of 60,000 cpm/well were added, with 0.072 IU (or 2 IU/mL) of unlabeled insulin (Actrapid; Novo Nordisk) added to the last two wells. The plates were incubated and examined under conditions similar to those described for the noncompetitive method. IAA levels were calculated as relative units and were related to positive controls. Positivity for IAA was set to 1.9 relative units. The competitive method was used to verify false-positive binding in the noncompetitive assay. In subsequent analysis, the competitive assay was used.

#### **Autoantibodies to ZnT8**

ZnT8 autoantibodies (ZnT8A) were analyzed in 5  $\mu$ L of serum with RBA, as described previously. (30) Duplicate samples were incubated with equal amounts of the three radio-labelled ZnT8 R/W/Q variants. Every putative positive sample (cut-off >59 U/mL) was analyzed for each ZnT8 variant separately.

#### **DIPP:**

The islet autoantibodies were analyzed in the Research Laboratory, Department of Pediatrics, University of Oulu, Oulu, Finland except for ZnT8A, which were analyzed in the PEDIA laboratory, University of Helsinki. The biochemical autoantibodies IAA, GADA, IA-2A, and ZnT8A were analyzed with specific radiobinding assays. Classical islet cell antibodies (ICA) were measured by indirect immunofluorescence and ICA was used as the primary screening tool for the beta-cell autoimmunity in children born in 1994-2002. IAA, IA-2A and GADA were measured from all samples obtained from children who became ICA positive. DIPP children born 2003 or later were monitored for ICA, IAA, IA-2A and GADA at every visit. ZnT8A were measured from all samples of children who seroconverted to positivity for at least one other biochemical islet autoantibody (IAA, IA-2A, GADA). In addition, all five islet autoantibodies were measured in the first 1004 children followed in the DIPP study. The reference values for the IAA, GADA, and IA-2A assays were based on the 99<sup>th</sup> percentile of more than 370 non-diabetic Finnish children and adolescents.

#### **Islet cell antibodies (ICA)**

ICA were quantified by standard immunofluorescence on sections of frozen human pancreas, blood group O donor. Fluorescein-conjugated antihuman IgG (Sigma-Aldrich Corp., St. Louis, MO) was used for detection. The end-point dilution titers of the ICA-positive samples were recorded, and the results were expressed in Juvenile Diabetes Foundation units (JDF-U). The detection limit was 2.5 JDF-U.

#### **Insulin Autoantibodies (IAA)**

Serum levels of IAA were quantified with a microassay modified from the method described by Williams et al. (31,32) Protein A-Sepharose (Pharmacia Biotech, Uppsala, Sweden) was used for precipitation of antibody-antigen complexes after incubation of the serum samples with mono- [125I]TyrA14-human insulin (Amersham Biosciences, Little Chalfont, UK) for 72 h in the absence or presence of an excess of unlabeled insulin. The IAA titers representing specific binding were expressed in relative units (RU) based on a standard curve run on each plate. A subject was considered positive for IAA when the specific binding exceeded 99th percentile in 371 nondiabetic Finnish subjects.

#### **Autoantibodies to GAD65 (GADA)**

GADA were measured by a radiobinding assay as previously described (33). The results were expressed in relative units on the basis of a standard curve. The cut-off limit for positivity was at the 99th percentile for 373 nondiabetic Finnish children and adolescents.

#### **Autoantibodies to islet antigen 2 (IA-2A)**

IA-2A were quantified by a radiobinding assay as previously described by (34). Antibody titers were expressed in relative units based on a standard curve. The cut-off limit for positivity was defined by the 99th percentile in 374 healthy Finnish children and adolescents.

#### **Autoantibodies to zinc transporter 8 (ZnT8A)**

ZnT8A were analyzed by a radiobinding assay modified from the method by Wenzlau et al. (35) and Salonen et al (36). A35S-methionine-labelled chimeric recombinant plasmid (4.1) encoding the COOH-terminal region (aa268–369) of the ZnT8 aa325-Arg allele and the ZnT8 aa325-Trp allele was used. The detection limit was set based on the levels of 250 nondiabetic Finnish children and adolescents.

Based on the results from the Diabetes Autoantibody Standardization Program (DASP) and the Islet Autoantibody Standardization Program (IASP) in 2010–2016 (19,21,22), the disease sensitivities of the IAA, GADA, IA-2A, and ZnT8A radiobinding assays have been 36–62%, 64–88%, 62–72%, and 62–70, respectively, while the corresponding disease specificities have been 94–98%, 94–99%, 93–100%, and 99–100%, respectively. ICA assay sensitivity in 2016 IASP workshop was 84% and specificity 88%.



## Technical Appendix

### Foundational features: (Table S1)

There are nine static variables: **1) Sex:** male and female, **2) Race:** black, white, other, biracial **3) Ethnicity:** Hispanic or non-Hispanic, **4) Birth Month,** **5) Birth Year.** Subjects' day of birth was not shared in compliance with the GDPR/HIPPA rules and all dates were shifted (transformed) by individual sites according to their local policy. **6) Type 1 diabetes Family History Status:** as yes/no **7) First Degree relationship** (harmonized to Mother, Father, Sibling, Multiple) **8) Breastfeeding ever status:** as yes/no **9) HLA Genotype (class II) markers - DRB, DQA1, DQB1.**

There are 9 dynamic variables: **1) Clinical Age:** in Years (resolution to 1 decimal point) for each visit (including diagnosis visit) **2) Height** (harmonized to centimeters), **3) Weight** (harmonized to Kilograms) **4) HbA1c** (harmonized to NGSP, US standard), **5) Random Glucose or Plasma Glucose,** **6) OGTT\_0** (harmonized to mg/dL), **7) OGTT\_120** (harmonized to mg/dL) **8) Islet autoantibody status:** as positive/negative for IAA – Insulin autoantibodies, GADA – GAD autoantibodies, IA2 – Insulinoma-associated antigen-2 antibodies. ZnT8 – Zinc transporter 8 autoantibodies (where available, these were measured after the study period from available samples if any only if one of the other three autoantibodies were positive) **9) Type 1 Diabetes Diagnosis Status:** as yes/no.

### Harmonized dataset: (Table S2)

The ratio of female to male subjects are similar in each cohort, all cohorts have a majority of male subjects. However, each cohort differed in race/ethnicity distributions (due to geographic differences) as well as in many other aspects. The European sites (DIPP, DiPiS and BABYDIAB) are homogeneous in racial/ ethnicity distributions (data not shown) while the DAISY and DEW-IT (US sites) cohorts are racially diverse. First-degree relatives accounted for all of BABYDIAB participants - 57% and 38% had, respectively, affected mother or father. Within US sites, 44% of DAISY subjects are first-degree relatives – 13%, 16%, and 14% have, respectively affected mother, father or sibling. Only 7% of DiPiS and 9% of DEW-IT participants have an affected relative.

### HLA risk groups in the harmonized dataset: (Table S3-S5)

Please refer to supplemental Table S5 for following details. In the harmonized dataset, 3368 (14%) subjects belong to group A, 11224 (46%) to group B, 3969 (16%) to group C, and 5847 (24%) to group D. There are 255 (~1%) of subjects who are not assigned an HLA group because of missing genotype (one haplotype or both missing and a missing allele). The proportion of subjects in the highest harmonized risk group (Group A) are similar in two US sites - DAISY, and DEW-IT (20%, 17% respectively) and in two EU sites - DiPiS and DIPP (12% and 14% respectively) but much lower (6%) in the BABYDIAB (Germany) site. Similarly, Group B proportion is similar in the two US sites - DAISY and DEWIT (47%, 45% respectively) but varies among the EU sites DIPP (66%), BABYDIAB (20%), DiPiS (5%). Overall Group C subject proportions in DIPP and DiPiS are smaller

than in other cohorts (12% each) vs. 20% in DAISY, 21% in BABYDIAB and 26% in DEW-IT. Lastly, among the EU sites, very few subjects in DIPP (8%) are in the lowest risk group, Group D, whereas for DiPiS they are the majority (71%) and sizeable for BABYDIAB (46%). For the US sites, both DAISY and DEW-IT have similar but smaller proportions (13% and 11% respectively) of Group D subjects. In the BABYDIAB cohort 6% of subjects remain unassigned to any group due to incomplete or missing HLA genotype.

### **Islet autoimmunity and risk of type 1 diabetes in following cohorts**

#### **A) Individual study cohorts: (Supplemental Figures S2(a-e))**

**BABYDIAB:** After 15 years follow up from confirmed seroconversion, type 1 diabetes developed in 38% (95% CI 22-59%) of subjects with single autoantibody, in 78% (58-92%) of subjects with two autoantibodies, and 84% (68-95%) of subjects with at least three autoantibodies at seroconversion respectively (Supplemental Figure 1a) with overall  $P < 0.0001$ .

**DAISY:** After 15 years follow up from confirmed seroconversion, type 1 diabetes developed in 39% (95% CI 28-50%) of subjects with single autoantibody, 76% (58-85%) of subjects with two autoantibodies, and 92% (82-95%) of subjects with at least three autoantibodies at seroconversion respectively (Supplemental Figure 1b) with overall  $P < 0.0001$ .

**DEW-IT:** After ~15 years follow up from confirmed seroconversion, type 1 diabetes had developed in 22% (95% CI 12-40%) of subjects with single autoantibody, 68% (48-85%) of subjects with two autoantibodies, and 75% (52-92%) of subjects with at least three autoantibodies at seroconversion by 15 years respectively (Supplemental Figure 1c) with overall  $P < 0.0001$ .

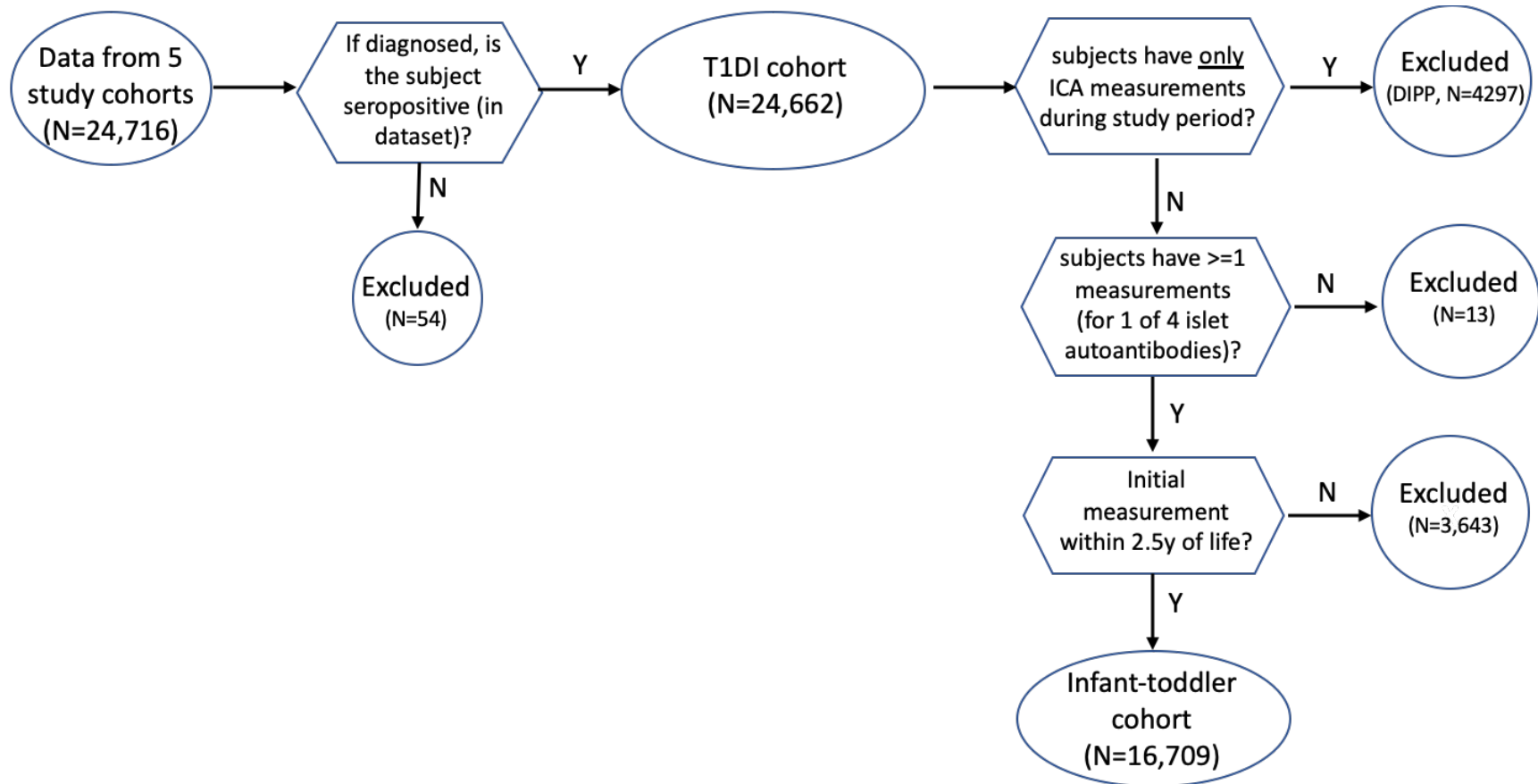
**DiPiS:** Since DiPiS started in early 2000s and are still following up subjects at age > 12, we report the incidence in the maximum follow up period for which we have data by number of autoantibodies at confirmed seroconversion. After follow-up of 10 years from seroconversion, type 1 diabetes had developed in 12% (95% CI 5-25%) of subjects with single autoantibody, 61% (35-85%) of subjects with two autoantibodies and 82% (65-92%) of subjects with at least three autoantibodies as these data were unavailable for computation of the incidence at 15 years (Supplemental Figure 1d) with overall  $P < 0.0001$ .

**DIPP:** After 15 years follow up from confirmed seroconversion, type 1 diabetes had developed in 45% (95% CI 38-50%) of subjects with single autoantibody, in 85% (75-91%) of subjects with two autoantibodies, and 91% (80-95%) of subjects with at least three autoantibodies by ~15 years as these data were unavailable for computation of the incidence at 15 years (Supplemental Figure 1e) with overall  $P < 0.0001$ .

**B) Harmonized T1DI cohort (overall cohort):** (Supplemental Figure S2(f))

After 15 years follow up from confirmed seroconversion, type 1 diabetes had developed in 40% (95% CI 35-45%) of subjects with single autoantibody, in 82% (75-88%) of subjects with two autoantibodies, and 90% (84-95%) of subjects with at least three autoantibodies (Supplemental Figure 2) with overall  $P < 0.0001$ .

**Supplemental Figure S1 - Derivation of the infant-toddler cohort**



**Supplemental Table S1 – Data variables submitted by individual study cohorts**

	Static variables – Subject-level*									Dynamic variables - Visit-level (follow up) †																
	Socio-Demo			Type1 Diabetes Family History		Diet	Genetic		Age at Diagno-sis	Growth		Glucose						Islet Autoantibodies†								Visit Age
												OGTT		A1c	FG	RG	GADA		IA2A		IAA		ZnT8A			
	DO B m	DOB y	Sex	FDR	Reln	BF ever	HLA loci	HLA Risk group		H	W	0	120							L	O	L	O	L	O	L
BABYDIAB	✓	✓	✓	✓	✓	✓	DR DQA1 DQB1	✓	✓	✓	✓	✓	✓	✓	✓		✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
DAISY	✓	✓	✓	✓	✓	✓	DR	✓	✓	✓	✓	✓	✓	✓		✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
DEW-IT	✓	✓	✓	✓	✓		DR DQA1 DQB1	✓	✓	✓	✓						✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
DiPiS	✓		✓	✓	✓	✓	DQA1 DQB1	✓	✓	✓	✓	✓	✓	✓		✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
DIPP	✓	✓	✓				DR DQA1 DQB1	✓	✓	✓	✓						✓	✓	✓	✓	✓	✓	✓	✓	✓	✓

\* FDR: First Degree Relative of Type 1 diabetes, ReIn: Relationship if any, BF ever: Breastfed ever (yes / no), H: Height, W: Weight

† L: Titer level of islet autoantibodies (*not harmonized as of this study*), O: Binary output (0 or 1), FG: Fasting Glucose, RG: Random Glucose

**Supplemental Table S2 – Characteristics of T1DI study cohort\*†**

Cohort	Sex (F)	Race				Hispanic	Breast fed ever (Y)	Fam Hx (Y) %, [Unknown Relationship n = ]	FDR (Relationship) N %			
		W	B	O	Biracial				Father	Mother	Sibling or half- sib	Multiple
<b>BABYDIAB (N=2376)</b>	1158 49%						1787 75%	2376 100%  [none]	895 38%	1344 57%	57 2%	80 3%
<b>DAISY (N=2544)</b>	1220 48%	2296 90%	62 2%	27 1%	139 6%	544 21%	2077 82%	1126 44%  [n = 1]	414 16%	324 13%	362 14%	25 1%
<b>DEW-IT (N=3756)</b>	1803 48%	2713 72%	15 <1%	25 <1%	149 4%	161 4%		355 10% [none]	132 4%	89 2%	81 2%	43 1%
<b>DiPiS (N=4360)</b>	2071 48%						3520 81%	308 7% [n = 103]	149 3%	129 3%	25 <1%	5 <1%
<b>DIPP (N=11680)</b>	5252 45%	11680 100%						not available				

\*All variables contain missing values, except Sex

† Contains all data, including for N=54 diagnosed cases with no follow up or positive islet autoantibodies during the study period which were excluded for all analyses

**Supplemental Table S3 – Haplotype groups\***

<b><u>DR3-DQ2.5 includes</u></b> (DR3)-DQA1*05-DQB1*02 (DR3)-DQA1*05:01-DQB1*02:01 DQ2 (Finland)		<b><u>DR4-DQ8.1 includes</u></b> DRB1*04:01-DQA1*03-DQB1*03:02 DRB1*04:02-DQA1*03-DQB1*03:02 DRB1*04:03-DQA1*03-DQB1*03:02 DRB1*04:04-DQA1*03-DQB1*03:02 DRB1*04:05-DQA1*03-DQB1*03:02 DRB1*04:08-DQA1*03-DQB1*03:02 DRB1*04:08-DQA1*03-DQB1*03:02 DRB1*04:13-DQA1*03-DQB1*03:02 (DR4)-DQA1*03:01-DQB1*03:02/4 (DR4)-DQA1*03-DQB1*03:02 (DR4/9)-DQA1*03:01-DQB1*03:02/3/4 DRB1*04:01-DQA1*03:01-DQB1*03:02 DRB1*04:04-DQA1*03:01-DQB1*03:02 DQA1*03:01-DQB1*03:02/3/4 (DR4)-DQA1*03:01-DQB1*03:02 (DR4)-DQA1*03:01-DQB1*03:02/4 (DR4)-DQA1*03-DQB1*03:02	
<b><u>X (Neutral) includes</u></b>	<b><u>Y (Protective) includes</u></b>	<b><u>Z (Highly Protective) includes</u></b>	
(DR1)-DQA1*01:01-DQB1*05:01	(DR11/12/13)-DQA1*05-DQB1*03:01	(DR13/14/15/16)-DQA1*01-DQB1*06	
(DR1/10)-DQA1*01-DQB1*05:01	(DR4)-DQA1*03-DQB1*03:01	(DR13)-DQA1*01:02-DQB1*06:09	
(DR1/10)-DQA1*01-DQB1*05:01	(DR4)-DQA1*03-DQB1*03:01	(DR13)-DQA1*01-DQB1*06:09	
(DR1/10)-DQB1*05:01	DRB1*04:01-DQA1*03-DQB1*03:01	(DR14)-DQA1*01:03-DQB1*05:03	
(DR4)-DQA1*030x-DQB1*04:01	DRB1*04:01/8-DQA1*03-DQB1*03:01	(DR14)-DQA1*01:04-DQB1*05:03	
(DR4)-DQA1*03:01-DQB1*04:01	DRB1*0408-DQA1*03-DQB1*03:01	(DR7)-DQA1*0201-DQB1*03:03	
(DR4)-DQA1*03:02-DQB1*02:02	(DR11/13)-DQA1*05-DQB1*03:01/9	(DR15)-DQB1*06:02	
(DR9)-DQA1*03:0x-DQB1*02:02	(DR11/13)-DQA1*05:01-DQB1*03:01	(DR13)-DQB1*06:03	
(DR9)-DQA1*03:02-DQB1*03:03	(DR11/13) -DQA1*05-DQB1*03:01/9	(DR13)-DQB1*06:03	
(DR9)-DQA1*03:01-DQB1*03:02	(DR11/13)-DQA1*05-DQB1*03:01	(DR13)-DQB1*06:09	
(DR9)-DQA1*03-DQB1*03:03	(DR15)-DQB1*06:01	(DR14)-DQB1*05:03	
(DR9)-DQA1*03:01-DQB1*03:04	(DR7)-DQA1*02:01-DQB1*02:02	(DR7)-DQA1*02:01-DQB1*03:03	
(DR9)-DQA1*03:01-DQB1*03:03	(DR7)-DQA1*02:01-DQB1*02:01	(DR15)-DQA1*01-DQB1*06:03	
(DR13)-DQA1*01:02-DQB1*06:04	(DR7)-DQA1*02:01-DQB1*02	(DR15)-DQA1*01-DQB1*06:02/3	

(DR13)-DQB1*06:04	(DR7)-DQA1*02:01-DQB1*03:01?	(DR15)-DQA1*01:03-DQB1*06:03
(DR16)-DQA1*01:02-DQB1*05:02	(DR4)-DQA1*03-DQB1*03:01	(DR15)-DQA1*01:03-DQB1*06:02/3
(DR16)-DQB1*05:02	(DR3)-DQA1*05:01-DQB1*03:01?	(DR15)-DQA1*01:02-DQB1*06:02
(DR8)-DQA1*04:01-DQB1*04:02	DRB1*0403-DQA1*03-DQB1*03:02	DRB1*0407-DQA1*03-DQB1*03:01
(DR8)-DQB1*04	(DR4)-DQA1*03:01-DQB1*03:01	
(DR13/14/15/16)-DQA1*01-DQB1*05/06	DRB1*0407-DQA1*03-DQB1*03:02	
	(DR4)-DQA1*03-DQB1*03:01	
	(DR8)-DQA1*03-DQB1*03:01	
	(DR15)-DQA1*01:03-DQB1*06:01	

\* Based on odds ratios from Table 2 (37)

**Supplemental Table S4 – HLA harmonization to 4 Risk Groups**

<u><b>Group A</b></u>	<u><b>Group B</b></u>	<u><b>Group C</b></u>	<u><b>Group D</b></u>
DR3-DQ2.5/DR4-DQ8.1	DR-DQ8.1/DR-DQ8.1 DR-DQ8.1/X DR3-DQ2.5/DR3-DQ2.5	DR-DQ8.1/Y DR3-DQ2.5/X DR-DQ8.1/X or Y or Z	DR3-DQ2.5/Y DR3-DQ2.5/Z DR-DQ8.1/Z X/Y Y/Y X/X X/Z Y/Z Z/Z  DR3-DQ2.5/X or Y



**Supplemental Table S5 – Summary of HLA risk group assignment**

<b>Cohort</b>	<b>Cohort Type</b>	<b>Cohort Subjects (N)</b>	<b>Group: A</b>	<b>Group: B</b>	<b>Group: C</b>	<b>Group: D</b>	<b>Unassigned*</b>
<b>BABYDIAB</b>	Overall	2376	134 (6%)	484 (20%)	510 (22%)	1097 (46%)	151 (6%)
	Infant / Toddler	2346	131 (6%)	477 (20%)	504 (22%)	1092 (47%)	142 (6%)
<b>DAISY</b>	Overall	2544	502 (20%)	1196 (47%)	501 (20%)	338 (13%)	7 (<1%)
	Infant / Toddler	2170	466 (22%)	1057 (49%)	410 (19%)	230 (11%)	7 (<1%)
<b>DEW-IT</b>	Overall	3756	627 (17%)	1674 (45%)	994 (27%)	407 (11%)	54 (1.4%)
	Infant / Toddler	559	157 (28%)	310 (56%)	37 (7%)	50 (9%)	5 (<1%)
<b>DiPiS</b>	Overall	4360	515 (12%)	199 (5%)	533 (12%)	3111 (71%)	2 (<1%)
	Infant / Toddler	4353	514 (12%)	199 (5%)	532 (12%)	3106 (71%)	2 (<1%)
<b>DIPP</b>	Overall	11680	1603 (14%)	7694 (66%)	1440 (12%)	899 (8%)	44 (<1%)
	Infant / Toddler	7281	944 (13%)	4589 (63%)	1025 (14%)	701 (10%)	22 (<1%))
<b>Totals† (N=24,716)</b>	Overall	<b>24716</b>	<b>3381 (14%)</b>	<b>11247 (45%)</b>	<b>3978 (16%)</b>	<b>5852 (24%)</b>	<b>258 (1%)</b>
	Infant / Toddler	<b>16709</b>	<b>2212 (13%)</b>	<b>6632 (40%)</b>	<b>2508 (15%)</b>	<b>5179 (31%)</b>	<b>144 (1%)</b>

\*Genotyping missing or indeterminate, †Contains all data, including for N=54 diagnosed cases with no follow up or positive islet autoantibodies during the study period which were excluded for all analyses

**Supplemental Table S6 – Cumulative incidence of type 1 diabetes by HLA risk group and number of islet autoantibodies at seroconversion in overall cohort (N=1613)**

HLA Class II	Num of islet antibodies	T1D Incidence in 10 Years or as noted					
		Single (1) (95% CI) (N=1191)				Two (2) (95% CI)    (Fig S3b)  @Serocon version (N=319)	Three (3) (95% CI) (Fig S3c)  @Seroconv ersion (N=103)
		@Sero conversion (Fig S3a)	@Seroconversion + 2 y (N=1009) (Fig S4a)				
		1*	S-0‡  (N=335)	S-S†  (N=419)	S-M§  (N=255)		
DR4-DQ8/DR3-DQ2.5	A	51% (45-60%)	25% (5-48%)	40% (28-50%)	68% (55-75%)	72% (55-95%)	88% (75-95%)
DR4-DQ8/DR4-DQ8 DR4-DQ8/neutral (x) DR3-DQ2/DR3-DQ2	B	32% (28-38%)	5% (4-18%)	25% (15-30%)	55% (45-75%)	65% (50-85%)	92% (80-95%)
DR4-DQ8/protective (y) DR3-DQ2/neutral (x) DR4-DQ8/unknown	C	25% (15-35%)	5% (4-15%)	25% (5-30%)	95% (75-98%)	80% (50-90%)	60% (30-88%) <i>in 5y</i>

DR4-DQ8/highly protective (z) DR3-DQ2/protective (y) DR3-DQ2/highly protective (z) DR3-DQ2/unknown Neutral (x) /any except DR3 or 4 Protective (y) /any except DR3 or 4 Highly protective (z) /any except DR 3 or 4	<b>D</b>	12% (8-35%)	2% (0-10%)	2% (0-10%)	65% (45-85%)	70% (56-95%)	80% (55-95%)
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\* (Group A vs D, Group A vs B, Group A vs C, Group B vs D:  $P < 0.0001$ ), (Group B vs C:  $P = 0.04$ ), (Group C vs D:  $P = 0.02$ )

† (Group A vs D  $P < 0.0001$ ), (Group B vs C:  $P = 0.01$ , Group B vs D:  $P = 0.0001$ ) (Group A vs B:  $P = 0.048$ ) (Group C vs A:  $P = 0.0002$ )

‡ (Group A vs B:  $P = 0.0013$ , Group A vs C:  $P = 0.015$ , Group A vs D:  $P = 0.008$ )

§ (Group A vs B:  $P = 0.03$ ) (Group B vs C:  $P = 0.01$ )

|| (Group A vs B:  $P = 0.05$ )

S-0: single autoantibody at seroconversion with reversion to autoantibody negativity 2 years post seroconversion

S-S: single autoantibody at seroconversion and at 2 years post seroconversion

S-M: single autoantibody at seroconversion progressed to multiple autoantibodies 2 years post seroconversion

**Supplemental Table S7 – Risk of progression to type 1 diabetes among children with multiple islet autoantibodies (at seroconversion or thereafter) during follow-up in overall cohort (N=912)**

Quartiles of age distribution at development of multiple islet autoantibodies	N	Cumulative 5-year incidence of diabetes	Cumulative 10-year incidence of diabetes	Cumulative 15-year incidence of diabetes	Average annual incidence of diabetes over 10 years (per 100 per year)
<= 2.08 years	228	62% [55%, 68%]	83% [77%, 89%]	92% [87%, 95%]	17.8
>2.08 and <= 4.13 years	228	42% [36%, 49%]	74% [67%, 80%]	87% [78%, 92%]	13.4
>4.13 and <= 8.16 years	228	31% [24%, 37%]	63% [55%, 70%]	75% [66%, 82%]	9.7
>8.16 and <= 21.2 years	228	27% [20%, 33%]	48% [35%, 59%]	56% [39%, 68%]	6.4
Total	912				11.8

## Figure captions

**Supplemental Figure S2** – Cumulative incidence of type 1 diabetes by the number of positive islet autoantibodies at seroconversion in individual studies and the combined (overall) cohort.

- (a) BABYDIAB
- (b) DAISY
- (c) DEWIT
- (d) DiPiS
- (e) DIPP
- (f) Combined cohort

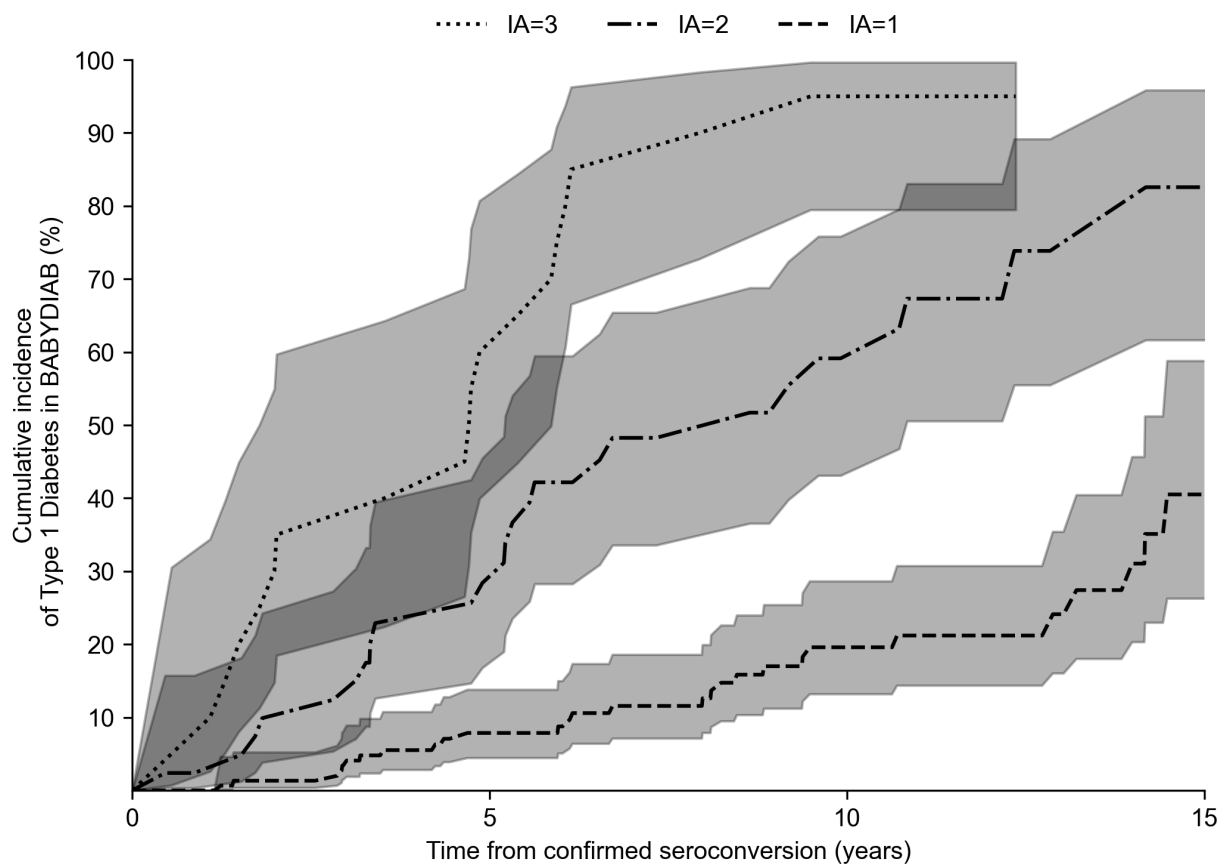
**Supplemental Figure S3** – Cumulative incidence of type 1 diabetes by the number of autoantibodies at seroconversion and HLA risk group in the combined (overall) cohort.

- (a) Single autoantibody
- (b) Two autoantibodies
- (c) Three autoantibodies

**Supplemental Figure S4** – Cumulative incidence of type 1 diabetes by HLA risk group among subjects with stable single autoantibody (S-S).

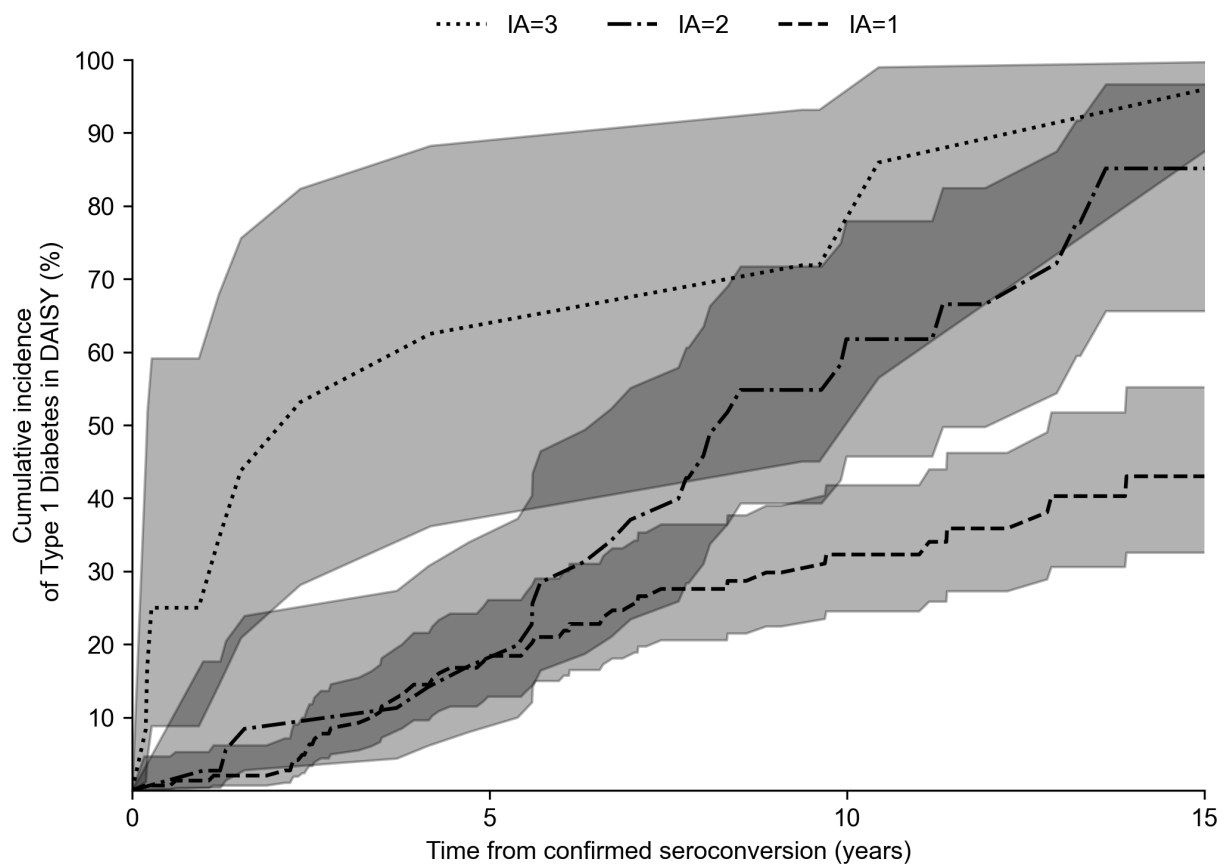
- (a) in the overall cohort
- (b) in the infant-toddler cohort

**Supplemental Figure S5** – Cumulative incidence of type 1 diabetes by age quartile at development of multiple positivity in the infant-toddler cohort.



Number at risk				
IA=3	20	8	1	0
IA=2	42	26	10	2
IA=1	158	113	56	8

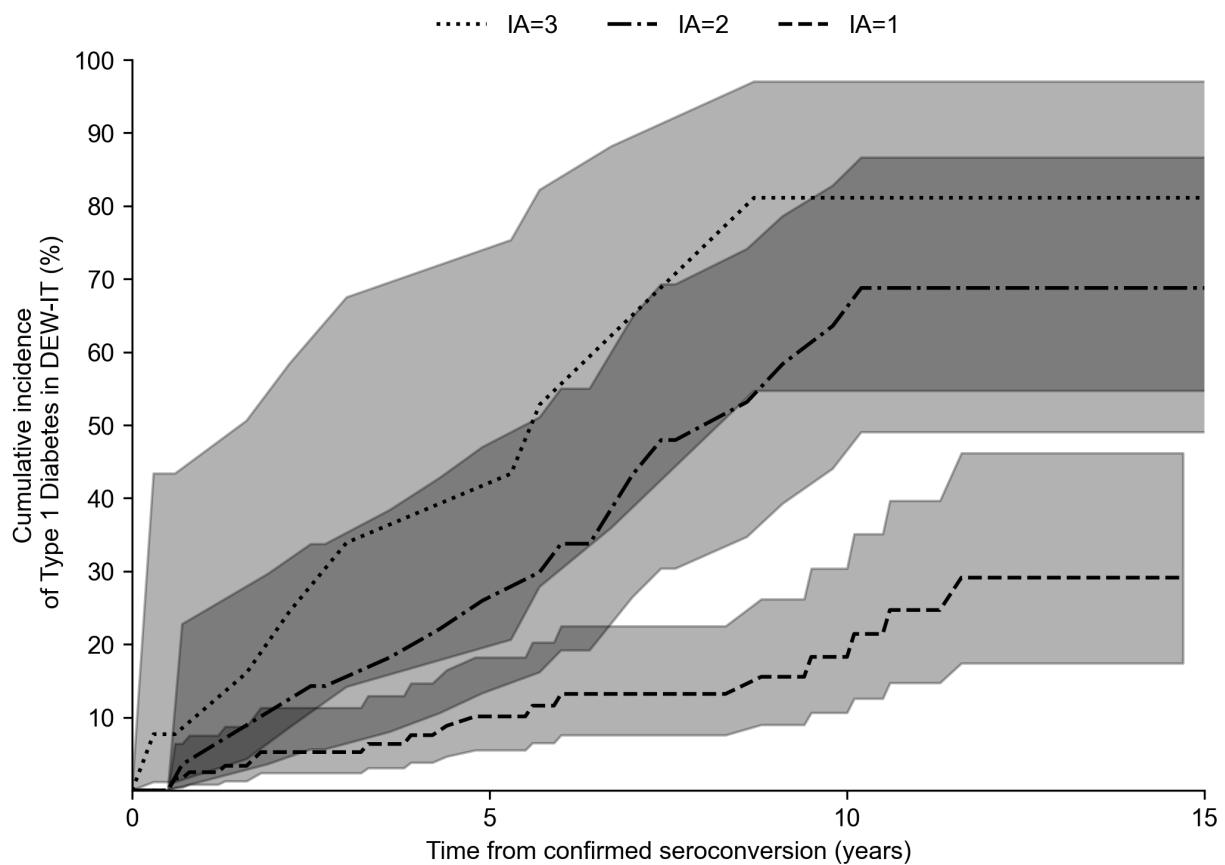
**Figure S2(a) - Cumulative incidence of type 1 diabetes by the number of positive islet autoantibodies at seroconversion in BABYDIAB (N=220 subjects, median age @ seroconversion: 5.57 years)**



Number at risk

IA=3	12	4	2	1
IA=2	37	29	11	1
IA=1	150	100	52	16

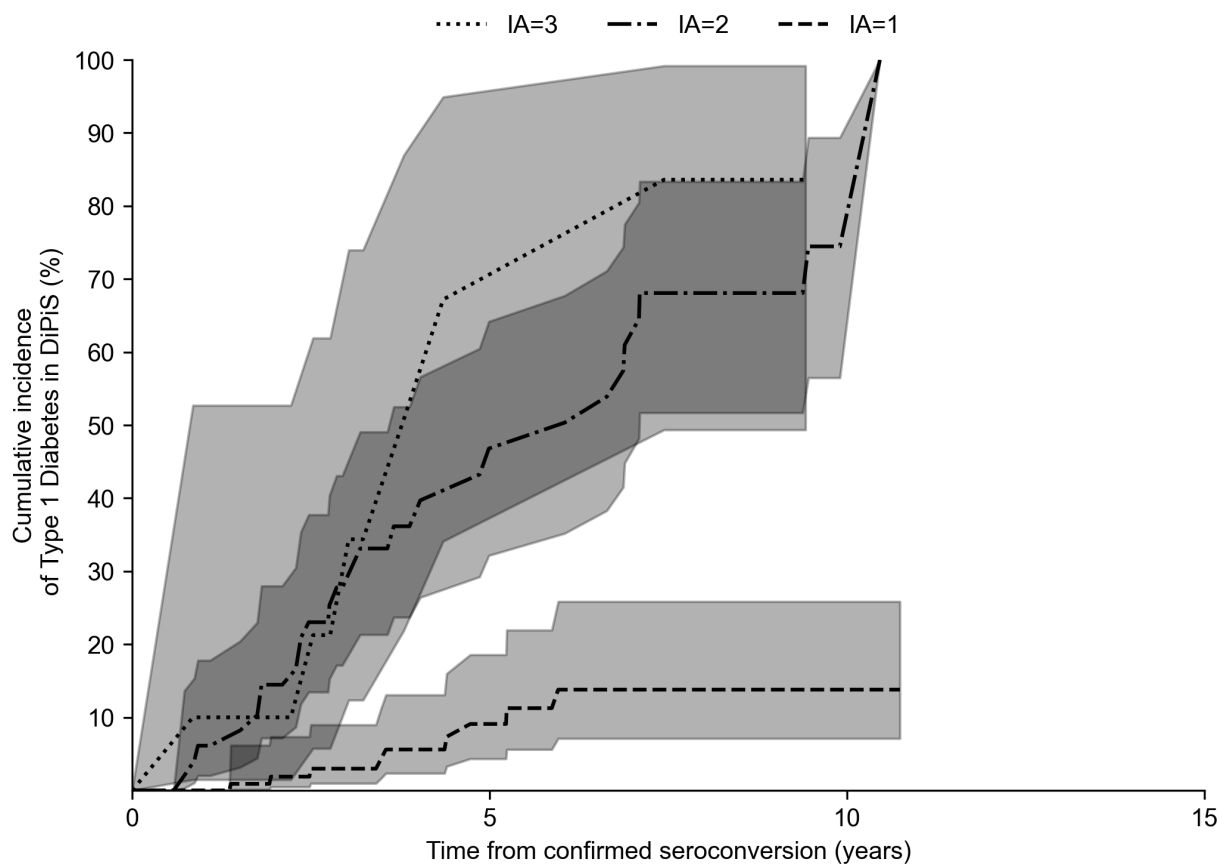
**Figure S2(b) - Cumulative incidence of type 1 diabetes by the number of positive islet autoantibodies at seroconversion in DAISY (N=199 subjects, median age @ seroconversion: 5.96 years)**



Number at risk				
IA=3	13	7	2	1
IA=2	32	19	7	1
IA=1	128	66	27	0

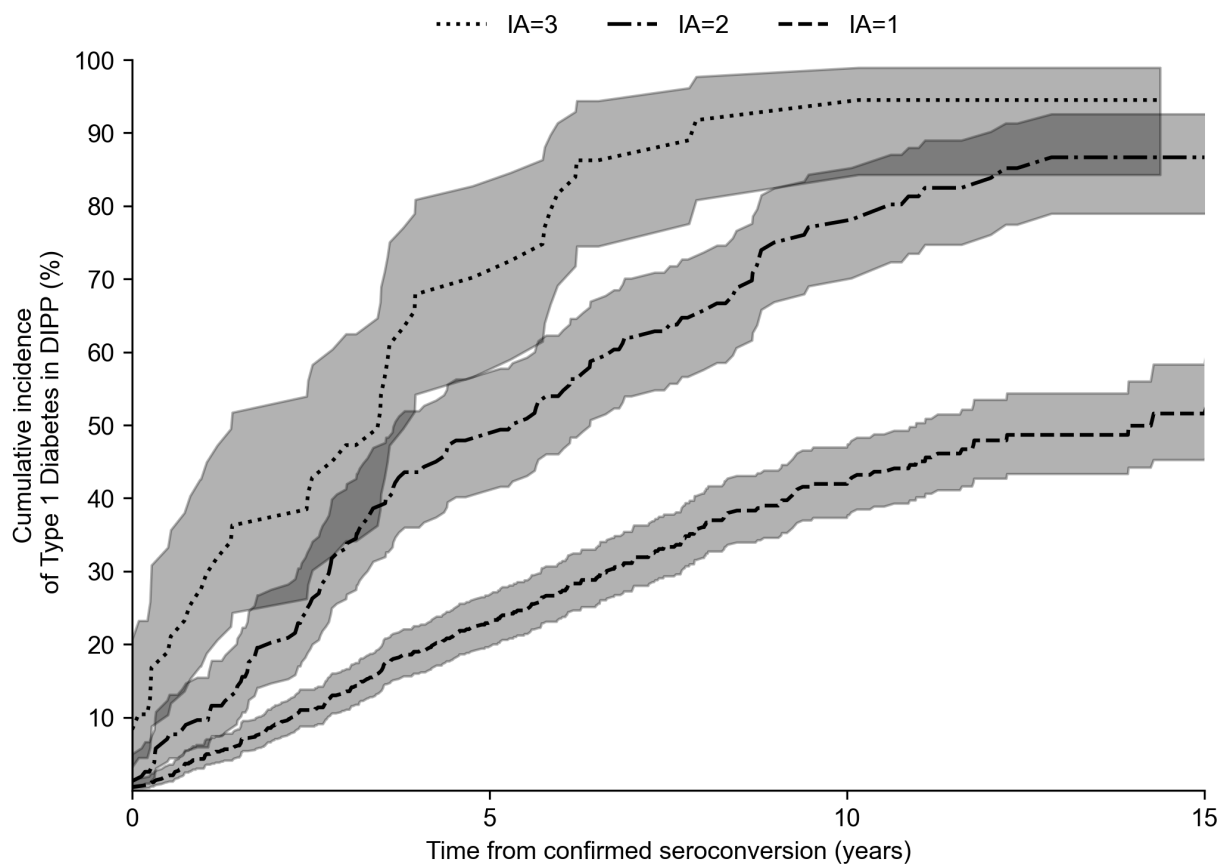
**Figure S2(c) - Cumulative incidence of type 1 diabetes by the number of positive islet autoantibodies at seroconversion in DEW-IT (N=173 subjects, median age @ seroconversion: 7.5 years)**





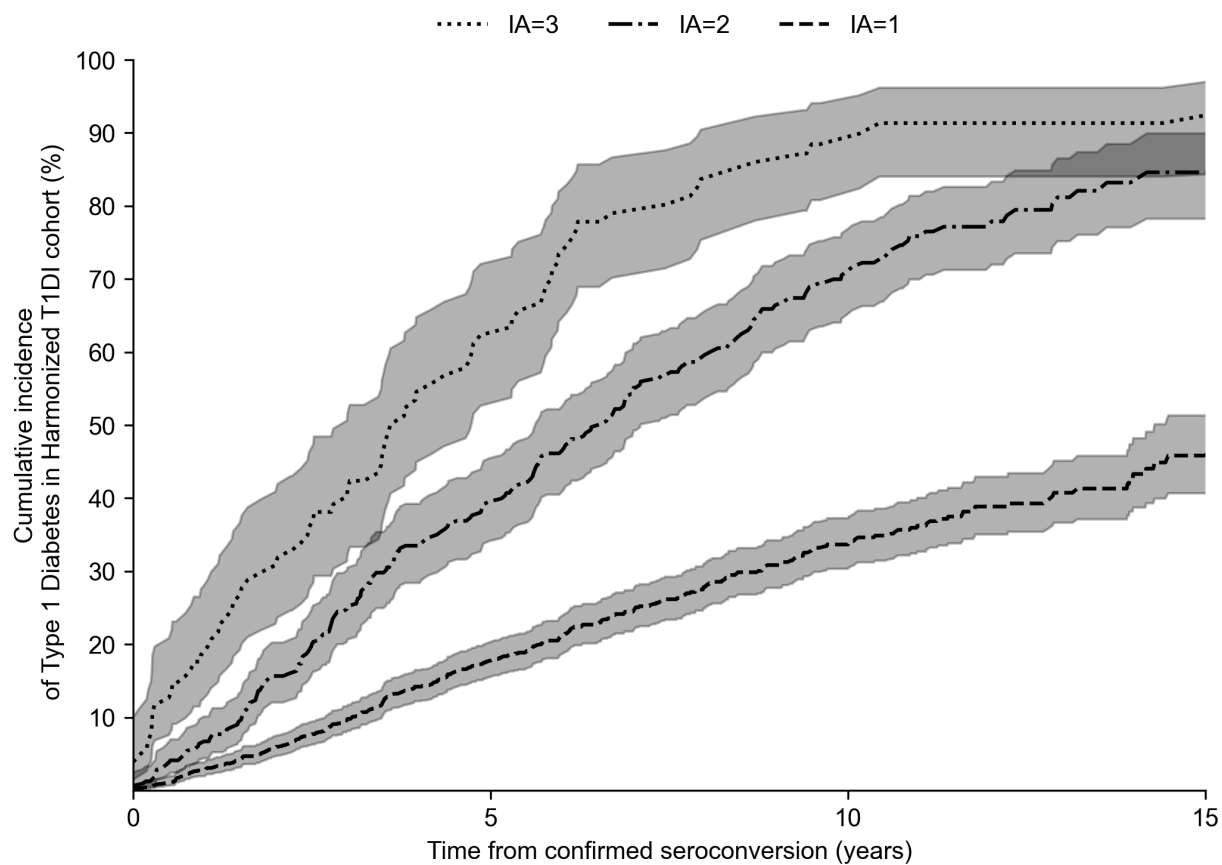
Number at risk				
IA=3	10	2	0	0
IA=2	51	15	1	0
IA=1	123	46	1	0

**Figure S2(d) - Cumulative incidence of type 1 diabetes by the number of positive islet autoantibodies at seroconversion in DiPiS (N=184 subjects, median age @ seroconversion: 7.03 years)**



Number at risk				
IA=3	48	13	3	0
IA=2	157	69	22	3
IA=1	632	360	144	19

**Figure S2(e) - Cumulative incidence of type 1 diabetes by the number of positive islet autoantibodies at seroconversion in DIPP (N=837 subjects, median age @ seroconversion: 3.04 years)**

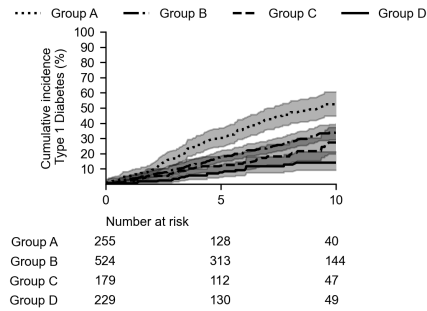


Number at risk

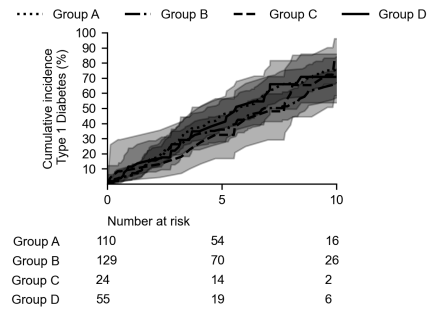
IA=3	103	34	8	2
IA=2	319	158	51	7
IA=1	1191	685	280	43

**Figure S2(f) - Cumulative incidence of type 1 diabetes by the number of positive islet autoantibodies at seroconversion in the overall cohort (N=1613 subjects, median age @ seroconversion: 4.6 year)**

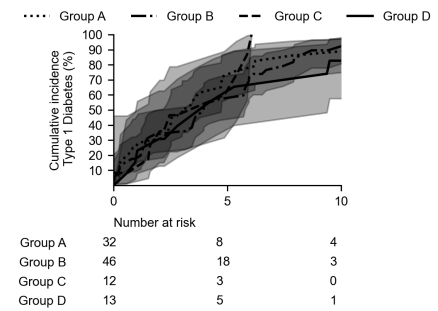
**Figure S3-1**



**Figure S3-1(a)**

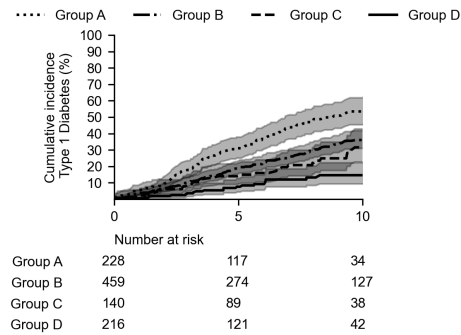


**Figure S3-1(b)**

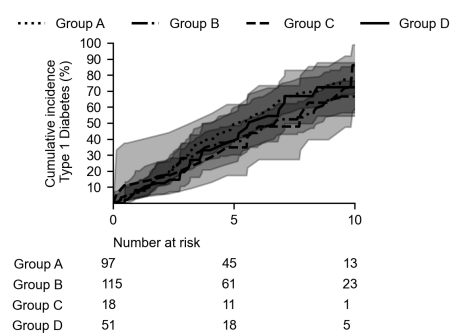


**Figure S3-1(c)**

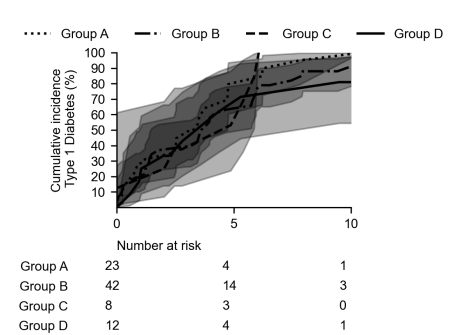
**Figure S3-2**



**Figure S3-2(a)**



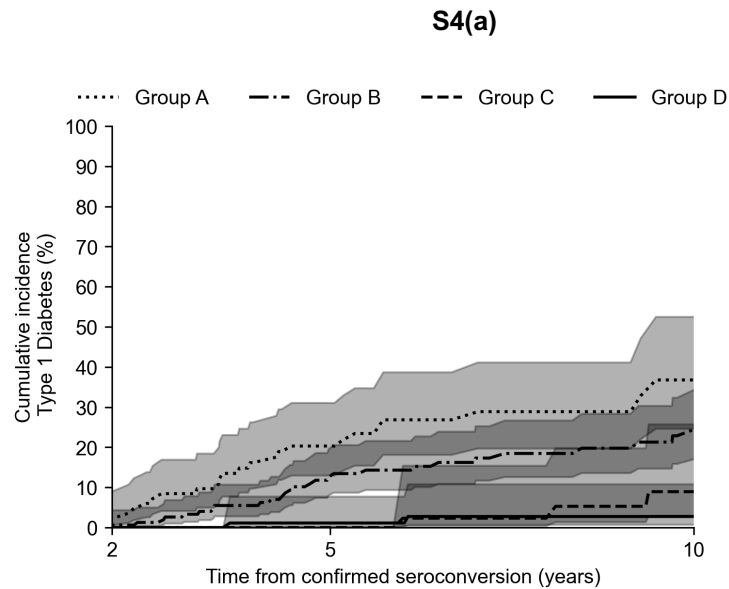
**Figure S3-2(b)**



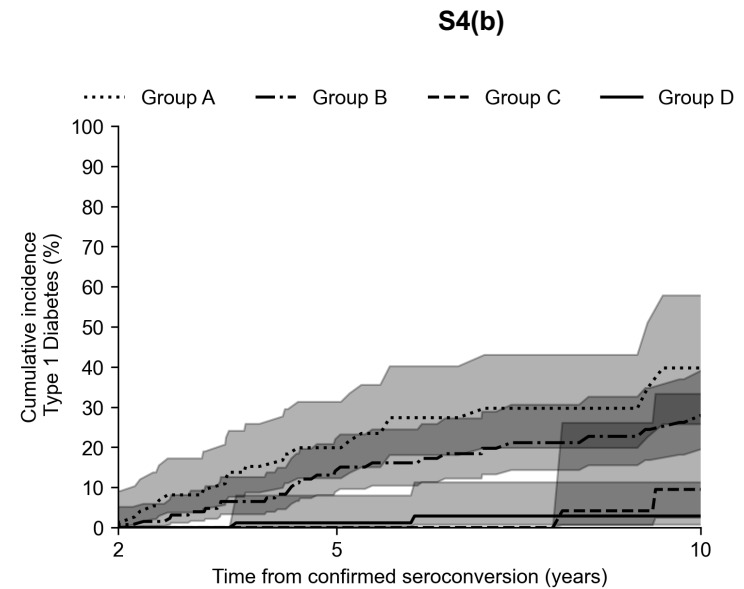
**Figure S3-2(c)**

Figures (a) – Single autoantibody, (b) – Two autoantibodies, (c) – Three autoantibodies

**Supplemental Figure S3 – Cumulative incidence of type 1 diabetes by the number of autoantibodies at seroconversion and HLA risk group in (3-1) - the overall cohort (N=1613) and in (3-2) - the infant-toddler cohort (N=1413).**



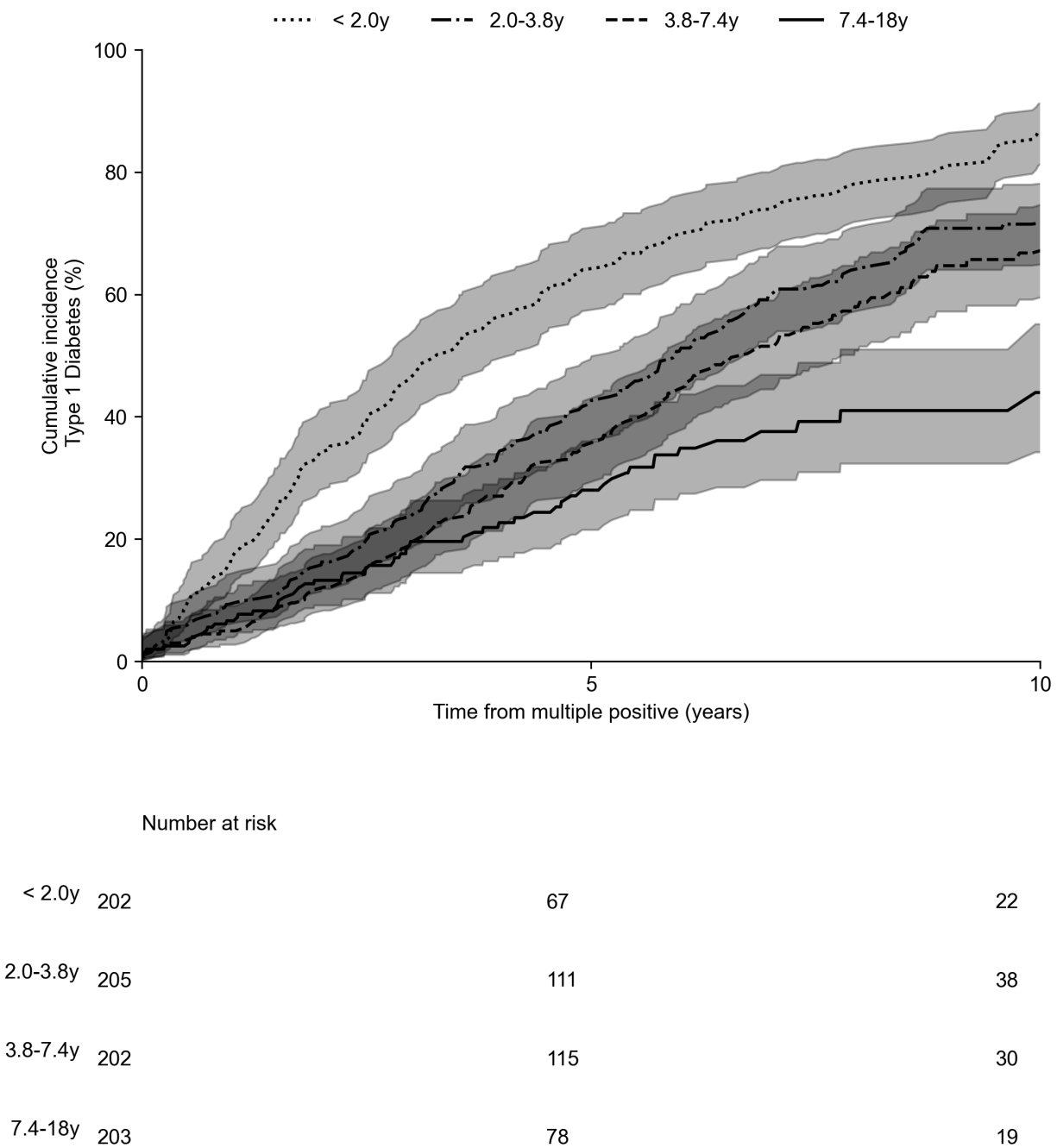
Number at risk			
Group A	82	52	15
Group B	161	106	48
Group C	63	50	21
Group D	108	70	24



Number at risk			
Group A	74	46	11
Group B	134	86	41
Group C	39	33	15
Group D	104	67	22

**Supplemental Figure S4 – Cumulative incidence of type 1 diabetes by HLA risk group among subjects with stable single autoantibody (S-S) in S4(a) - in the overall cohort and S4(b) - in the infant-toddler cohort.**

**Supplemental Figure S5 – Cumulative incidence of type 1 diabetes by age quartile at development of multiple positivity in the infant-toddler cohort.**



## References:

1. Ziegler AG, Hummel M, Schenker M, Bonifacio E. Autoantibody appearance and risk for development of childhood diabetes in offspring of parents with type 1 diabetes: the 2-year analysis of the German BABYDIAB Study. *Diabetes*. 1999 Mar;48(3):460–8.
2. Hummel S, Pflüger M, Hummel M, Bonifacio E, Ziegler A-G. Primary dietary intervention study to reduce the risk of islet autoimmunity in children at increased risk for type 1 diabetes: the BABYDIET study. *Diabetes Care*. 2011 Jun;34(6):1301–5.
3. Rewers M, Bugawan TL, Norris JM, Blair A, Beaty B, Hoffman M, et al. Newborn screening for HLA markers associated with IDDM: diabetes autoimmunity study in the young (DAISY). *Diabetologia*. 1996 Jul;39(7):807–12.
4. Rewers M, Norris JM, Eisenbarth GS, Erlich HA, Beaty B, Klingensmith G, et al. Beta-Cell Autoantibodies in Infants and Toddlers without IDDM Relatives: Diabetes Autoimmunity Study in the Young (DAISY). *J Autoimmun*. 1996 Jun;9(3):405–10.
5. Wion E, Brantley M, Stevens J, Gallinger S, Peng H, Glass M, et al. Population-wide infant screening for HLA-based type 1 diabetes risk via dried blood spots from the public health infrastructure. *Ann N Y Acad Sci*. 2003 Nov;1005:400–3.
6. Woo W, LaGasse JM, Zhou Z, Patel R, Palmer JP, Campus H, et al. A novel high-throughput method for accurate, rapid, and economical measurement of multiple type 1 diabetes autoantibodies. *J Immunol Methods*. 2000 Oct 20;244(1–2):91–103.
7. Larsson HE. A Swedish approach to the prevention of type 1 diabetes. *Pediatr Diabetes*. 2016 Jul 1;17(S22):73–7.
8. Larsson HE, Lynch K, Lernmark B, Nilsson A, Hansson G, Almgren P, et al. Diabetes-associated HLA genotypes affect birthweight in the general population. *Diabetologia*. 2005 Aug 1;48(8):1484–91.
9. Lundgren M, Sahlin Å, Svensson C, Carlsson A, Cedervall E, Jönsson B, et al. Reduced morbidity at diagnosis and improved glycemic control in children previously enrolled in DiPiS follow-up. *Pediatr Diabetes*. 2014 Nov;15(7):494–501.
10. Kupila A, Muona P, Simell T, Arvilommi P, Savolainen H, Hämäläinen AM, et al. Feasibility of genetic and immunological prediction of type I diabetes in a population-based birth cohort. *Diabetologia*. 2001 Mar;44(3):290–7.
11. Sjöroos M, Iltiä A, Ilonen J, Reijonen H, Lövgren T. Triple-label hybridization assay for type-1 diabetes-related HLA alleles. *BioTechniques*. 1995 May;18(5):870–7.

12. Hermann R, Turpeinen H, Laine AP, Veijola R, Knip M, Simell O, et al. HLA DR-DQ-encoded genetic determinants of childhood-onset type 1 diabetes in Finland: an analysis of 622 nuclear families. *Tissue Antigens*. 2003 Aug;62(2):162–9.
13. Ilonen J, Hammais A, Laine A-P, Lempainen J, Vaarala O, Veijola R, et al. Patterns of  $\beta$ -cell autoantibody appearance and genetic associations during the first years of life. *Diabetes*. 2013 Oct;62(10):3636–40.
14. Siljander HTA, Simell S, Hekkala A, Lähde J, Simell T, Vähäsalo P, et al. Predictive characteristics of diabetes-associated autoantibodies among children with HLA-conferred disease susceptibility in the general population. *Diabetes*. 2009 Dec;58(12):2835–42.
15. Parikka V, Näntö-Salonen K, Saarinen M, Simell T, Ilonen J, Hyöty H, et al. Early seroconversion and rapidly increasing autoantibody concentrations predict prepubertal manifestation of type 1 diabetes in children at genetic risk. *Diabetologia*. 2012 Jul;55(7):1926–36.
16. Achenbach P, Lampasona V, Landherr U, Koczwara K, Krause S, Grallert H, et al. Autoantibodies to zinc transporter 8 and SLC30A8 genotype stratify type 1 diabetes risk. *Diabetologia*. 2009 Sep;52(9):1881–8.
17. Bonifacio E, Yu L, Williams AK, Eisenbarth GS, Bingley PJ, Marcovina SM, et al. Harmonization of glutamic acid decarboxylase and islet antigen-2 autoantibody assays for national institute of diabetes and digestive and kidney diseases consortia. *J Clin Endocrinol Metab*. 2010 Jul;95(7):3360–7.
18. Williams AJ, Bingley PJ, Bonifacio E, Palmer JP, Gale EA. A novel micro-assay for insulin autoantibodies. *J Autoimmun*. 1997 Oct;10(5):473–8.
19. Törn C, Mueller PW, Schlosser M, Bonifacio E, Bingley PJ, Participating Laboratories. Diabetes Antibody Standardization Program: evaluation of assays for autoantibodies to glutamic acid decarboxylase and islet antigen-2. *Diabetologia*. 2008 May;51(5):846–52.
20. Wyatt R, Williams AJK. Islet Autoantibody Analysis: Radioimmunoassays. *Methods Mol Biol Clifton NJ*. 2016;1433:57–83.
21. Lampasona V, Schlosser M, Mueller PW, Williams AJK, Wenzlau JM, Hutton JC, et al. Diabetes Antibody Standardization Program: First Proficiency Evaluation of Assays for Autoantibodies to Zinc Transporter 8. *Clin Chem*. 2011 Dec 1;57(12):1693–702.
22. Lampasona V, Pittman DL, Williams AJ, Achenbach P, Schlosser M, Akolkar B, et al. Islet Autoantibody Standardization Program 2018 Workshop: Interlaboratory Comparison of Glutamic Acid Decarboxylase Autoantibody Assay Performance. *Clin Chem*. 2019 Sep 1;65(9):1141–52.



23. Grubin CE, Daniels T, Toivola B, Landin-Olsson M, Hagopian WA, Li L, et al. A novel radioligand binding assay to determine diagnostic accuracy of isoform-specific glutamic acid decarboxylase antibodies in childhood IDDM. *Diabetologia*. 1994 Apr 1;37(4):344–50.
24. Yu L, Rewers M, Gianani R, Kawasaki E, Zhang Y, Verge C, et al. Antiislet autoantibodies usually develop sequentially rather than simultaneously. *J Clin Endocrinol Metab*. 1996 Dec;81(12):4264–7.
25. Gianani R, Rabin DU, Verge CF, Yu L, Babu SR, Pietropaolo M, et al. ICA512 autoantibody radioassay. *Diabetes*. 1995 Nov;44(11):1340–4.
26. Vardi P, Dib SA, Tuttleman M, Connelly JE, Grinbergs M, Radizabeh A, et al. Competitive insulin autoantibody assay. Prospective evaluation of subjects at high risk for development of type I diabetes mellitus. *Diabetes*. 1987 Nov;36(11):1286–91.
27. Yu L, Robles DT, Abiru N, Kaur P, Rewers M, Kelemen K, et al. Early expression of antiinsulin autoantibodies of humans and the NOD mouse: evidence for early determination of subsequent diabetes. *Proc Natl Acad Sci U S A*. 2000 Feb 15;97(4):1701–6.
28. Wenzlau JM, Juhl K, Yu L, Moua O, Sarkar SA, Gottlieb P, et al. The cation efflux transporter ZnT8 (Slc30A8) is a major autoantigen in human type 1 diabetes. *Proc Natl Acad Sci U S A*. 2007 Oct 23;104(43):17040–5.
29. Tiberti C, Yu L, Lucantoni F, Panimolle F, Spagnuolo I, Lenzi A, et al. Detection of four diabetes specific autoantibodies in a single radioimmunoassay: an innovative high-throughput approach for autoimmune diabetes screening. *Clin Exp Immunol*. 2011 Dec;166(3):317–24.
30. Vaziri-Sani F, Delli AJ, Elding-Larsson H, Lindblad B, Carlsson A, Forsander G, et al. A novel triple mix radiobinding assay for the three ZnT8 (ZnT8-RWQ) autoantibody variants in children with newly diagnosed diabetes. *J Immunol Methods*. 2011 Aug 31;371(1–2):25–37.
31. Williams AJ, Bingley PJ, Bonifacio E, Palmer JP, Gale EA. A novel micro-assay for insulin autoantibodies. *J Autoimmun*. 1997 Oct;10(5):473–8.
32. Kukko M, Kimpimäki T, Korhonen S, Kupila A, Simell S, Veijola R, et al. Dynamics of Diabetes-Associated Autoantibodies in Young Children with Human Leukocyte Antigen-Conferred Risk of Type 1 Diabetes Recruited from the General Population. *J Clin Endocrinol Metab*. 2005 May 1;90(5):2712–7.
33. Savola K, Sabbah E, Kulmala P, Vähäsalo P, Ilonen J, Knip M. Autoantibodies associated with Type I diabetes mellitus persist after diagnosis in children. *Diabetologia*. 1998 Nov;41(11):1293–7.

34. Savola K, Bonifacio E, Sabbah E, Kulmala P, Vähäsalo P, Karjalainen J, et al. IA-2 antibodies - a sensitive marker of IDDM with clinical onset in childhood and adolescence. *Diabetologia*. 1998 Mar 20;41(4):424–9.
35. Wenzlau JM, Liu Y, Yu L, Moua O, Fowler KT, Rangasamy S, et al. A common nonsynonymous single nucleotide polymorphism in the SLC30A8 gene determines ZnT8 autoantibody specificity in type 1 diabetes. *Diabetes*. 2008 Oct;57(10):2693–7.
36. Salonen KM, Ryhänen S, Härkönen T, Ilonen J, Knip M, Finnish Pediatric Diabetes Register. Autoantibodies against zinc transporter 8 are related to age, metabolic state and HLA DR genotype in children with newly diagnosed type 1 diabetes. *Diabetes Metab Res Rev*. 2013 Nov;29(8):646–54.
37. Erlich H, Valdes AM, Noble J, Carlson JA, Varney M, Concannon P, et al. HLA DR-DQ Haplotypes and Genotypes and Type 1 Diabetes Risk: Analysis of the Type 1 Diabetes Genetics Consortium Families. *Diabetes*. 2008 Apr 1;57(4):1084–92.