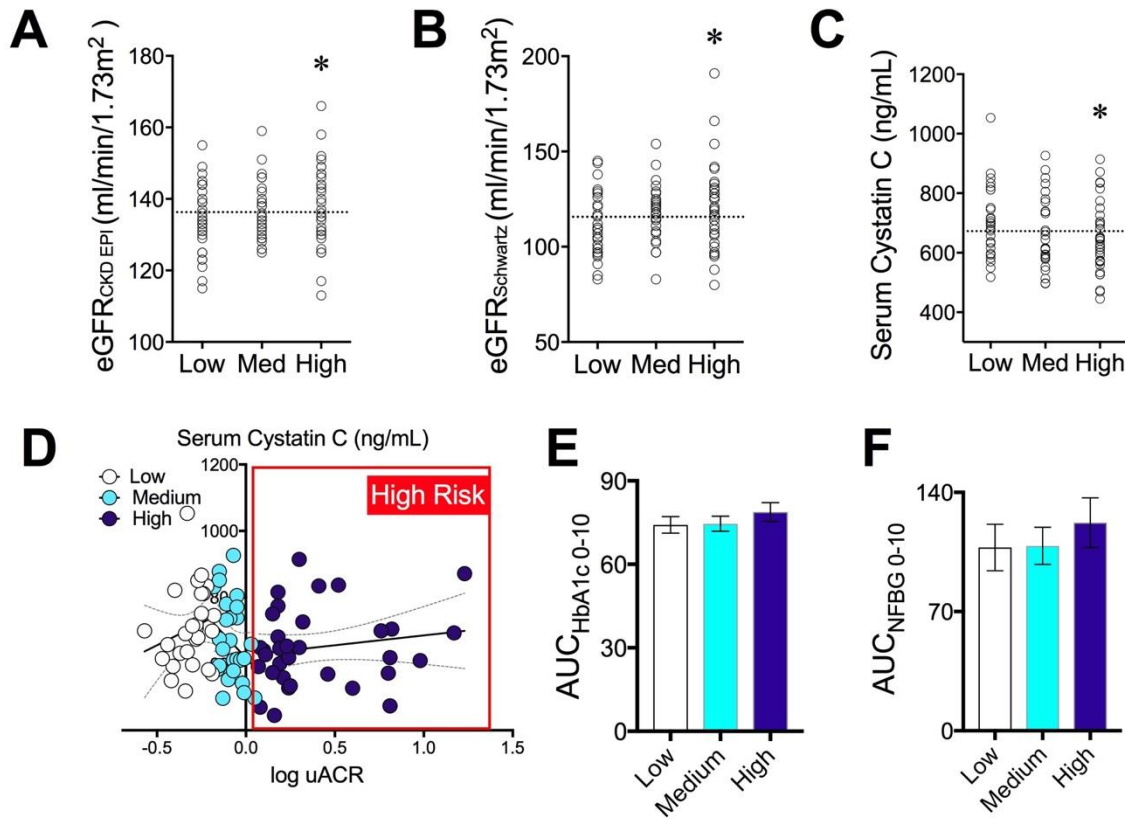


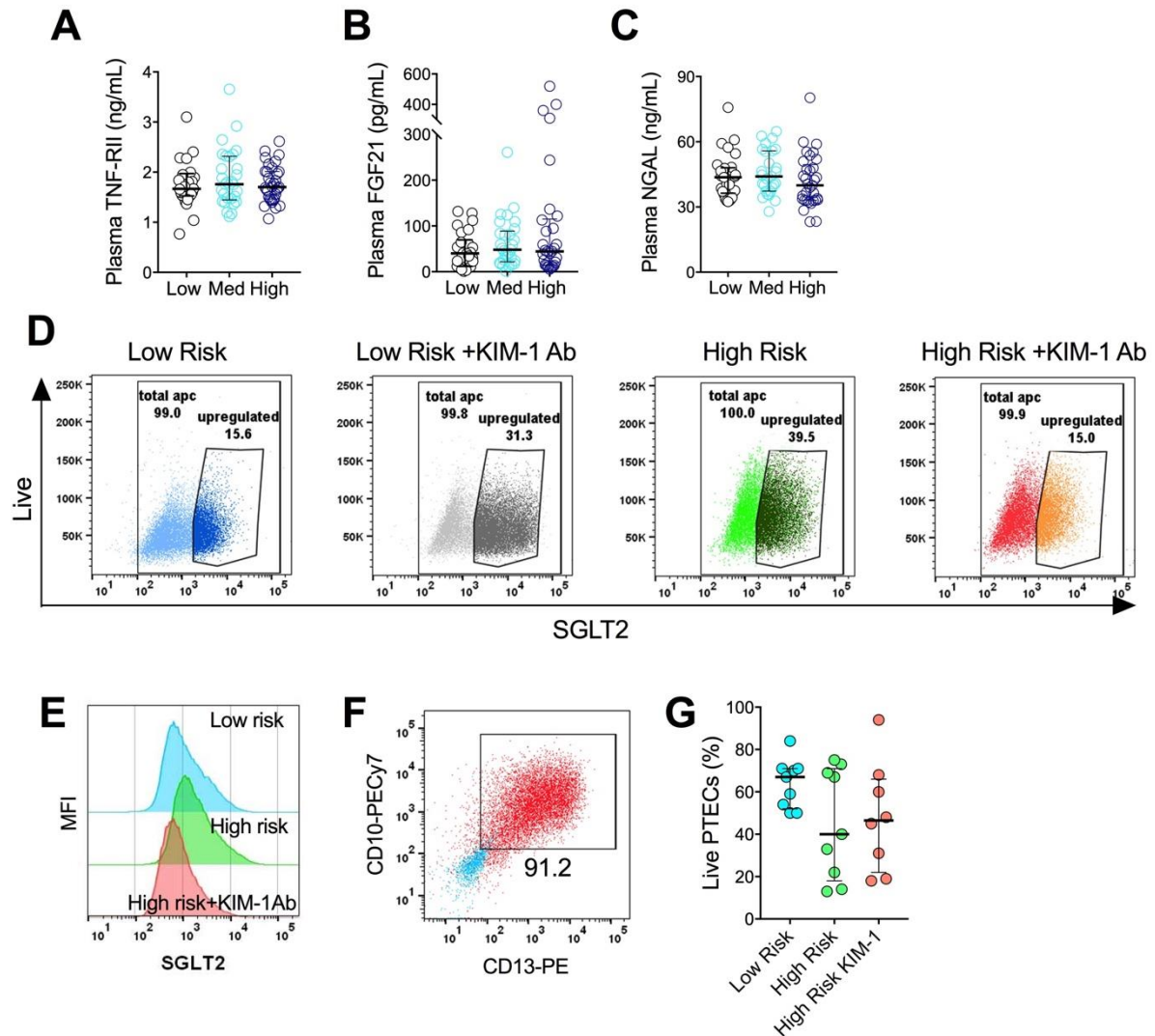
Forbes et al Supplementary Appendix; “T cell expression and release of kidney injury molecule-1 in response to glucose variations initiates kidney injury in early diabetes”:

Parameters	Control Healthy Tissue	Minimal Change Disease	1° FSGS	Diabetic Kidney Disease
Age (years)	50.3±22.4	23.3±5.9	54.5±20.3	51.5±17.9
Sex N, (% female)	4 (0)	4 (25)	4 (50)	4 (25)
BMI (kg/m ²)	N/A	N/A	N/A	N/A
Random BG (mmol/L)	6.4±1.2	5.1±1.0	6.4±2.2	9.6±0.4
Diabetes Duration (years)	0	No Family History	No Family History	11.8±6.7
HbA _{1C} (%;mmol/mol)	No record (available for one of four patients)	No record	No record (available for one of four patients)	8.7±4.0
SBP (mmHg)	No record	132.5±12.6	138.8±13.1	142.5±8.7
Total Chol (mmol/L)	No record (available for one of four patients)	No record (available for two of four patients)	5.1±0.4	5.4±1.1
eGFR _{CKD EPI} (ml/min/1.73m ²)	76.5±10.7	85.8±8.5	36.8±19.8	27.5±16.7
uPCR (g/mol)	No record (available for one of four patients)	686.3±640.3	244.3±171.7	684.8±491.5

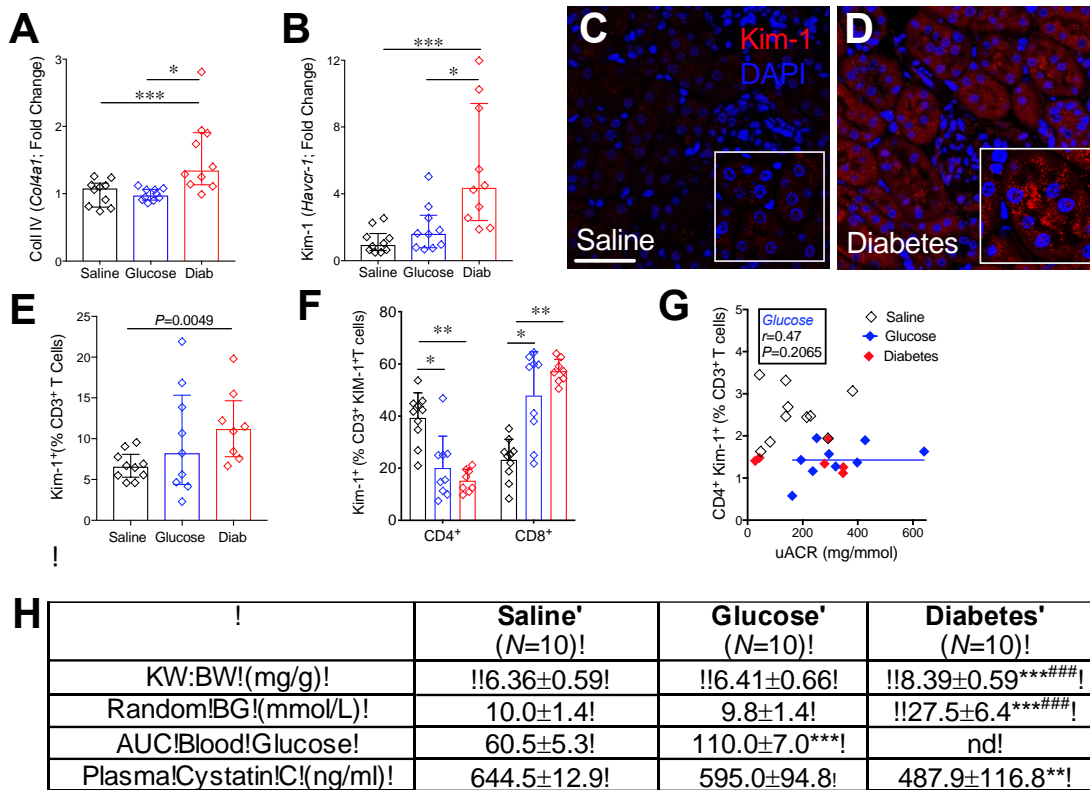
Suppl. Table S1. – Demographic and clinical characteristics of human kidney tissue donors. Control kidney tissue was obtained following Nephrectomy. Minimal Change Disease – immunofluorescence negative; IF/TA negative. Diabetic kidney disease – confirmed by the presence of Kimmelstiel-Wilson nodules, mesangial matrix expansion, immunofluorescence positive, IF/TA positivity. Focal Sclerosing Glomerulosclerosis (FSGS) – phenotypically not secondary FSGS; immunofluorescence negative; IF/TA positive. Minimal Change Disease – immunofluorescence negative; IF/TA negative. uPCR - urinary protein excretion/mol of creatinine. Data presented are Mean±SD.



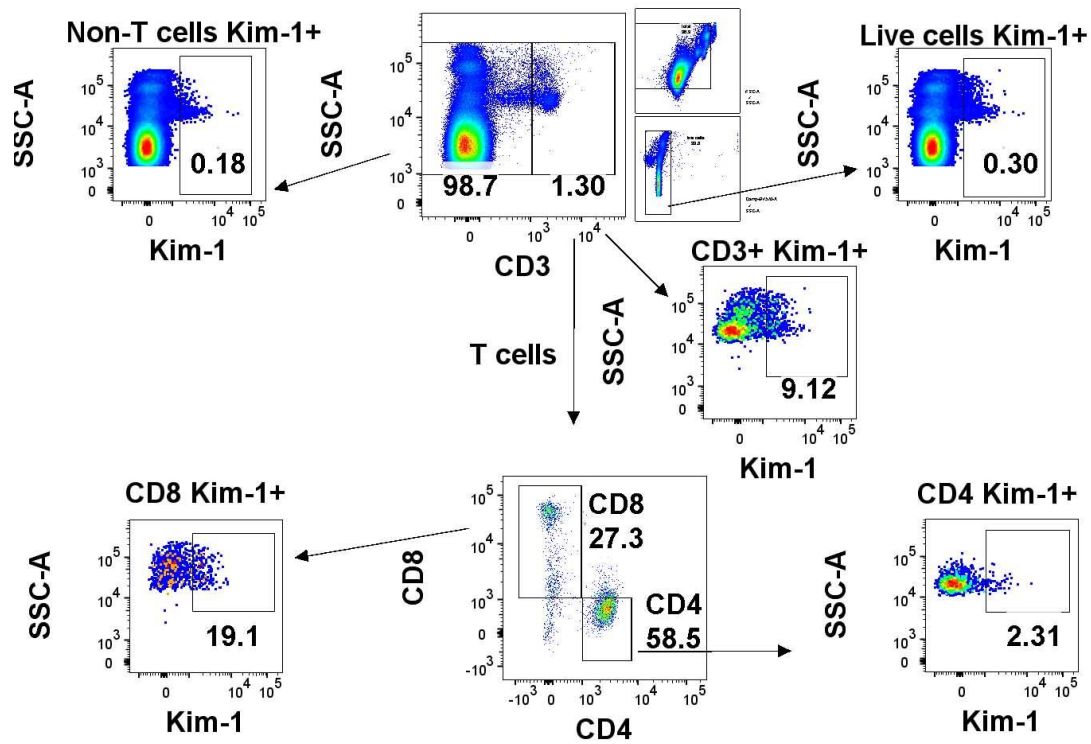
Suppl. Fig. S1. Kidney function data in youth with varied risk for DKD. Youth (20.0 ± 2.8 yrs old) with type 1 diabetes were stratified by risk for DKD using tertiles of urinary albumin:creatinine ratio (uACR; Low Risk, $N=33$; Medium Risk, $N=33$; High Risk, $N=34$). **A-C** Renal functional data plots of estimated glomerular filtration rate (eGFR) with the population mean shown as a dotted line using (A) the adult CKD-EPI equation; (B) the pediatric modified Schwartz equation; (C) the surrogate inverse marker serum cystatin C. (D) General linear regression plot of log uACR vs serum cystatin C corrected for age, sex, diabetes duration and HbA_{1c}. **E-F** Area under the curve (0-10 years) for historical (E) long term glucose control by HbA_{1c}; and (F) Random blood glucose concentrations (NFBG). * $P < 0.05$; ** $P < 0.01$ vs low risk group.



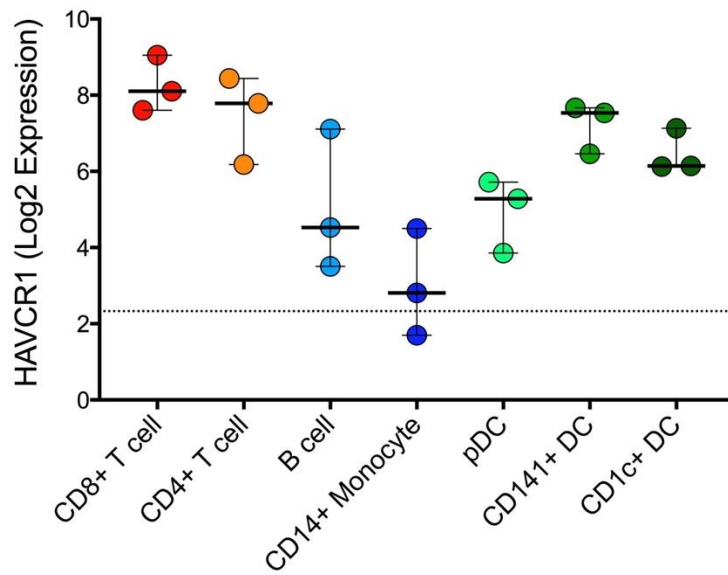
Suppl. Fig. S2. Biomarkers of kidney injury and proximal tubule cells exposed to plasma from youth at risk for DKD. Youth (20.0 ± 2.8 yrs old) with type 1 diabetes were stratified by risk for DKD using tertiles of urinary albumin:creatinine ratio (uACR; Low Risk , $N=33$; Medium Risk, $N=33$; High Risk, $N=34$). **A-C** Plasma biomarkers were measured by ELISA (**A**) tumor necrosis factor receptor -2 (TNF-RII); (**B**) fibroblast growth factor -21 (FGF-21) and (**C**) Neutrophil gelatinase-associated lipocalin (NGAL) in all youth within the cohort. **D-G** Human healthy primary proximal tubule epithelial cells (PTEC) were exposed to plasma (4%) from youth with varied risk for DKD for 24 hours in the presence and absence of KIM-1 blockade (Ab). (**D**) Representative Flow cytometry dot plots of shift in SGLT2-APC expression in PTEC following exposure to 4% patient plasma. (**E**) Representative flow cytometry dot plot characterizing live kidney cells as human PTEC by dual expression of CD10 and CD13 after exposure to patient plasma; (**F**) Proportion of live PTEC after 24 hours exposure to plasma; (**G**) Flow cytometry histograms of SGLT2 expression in live PTECs at 24 hours (blue-Low Risk; green-High Risk+Control Ab; red -High Risk+KIM-1 blocking Ab).



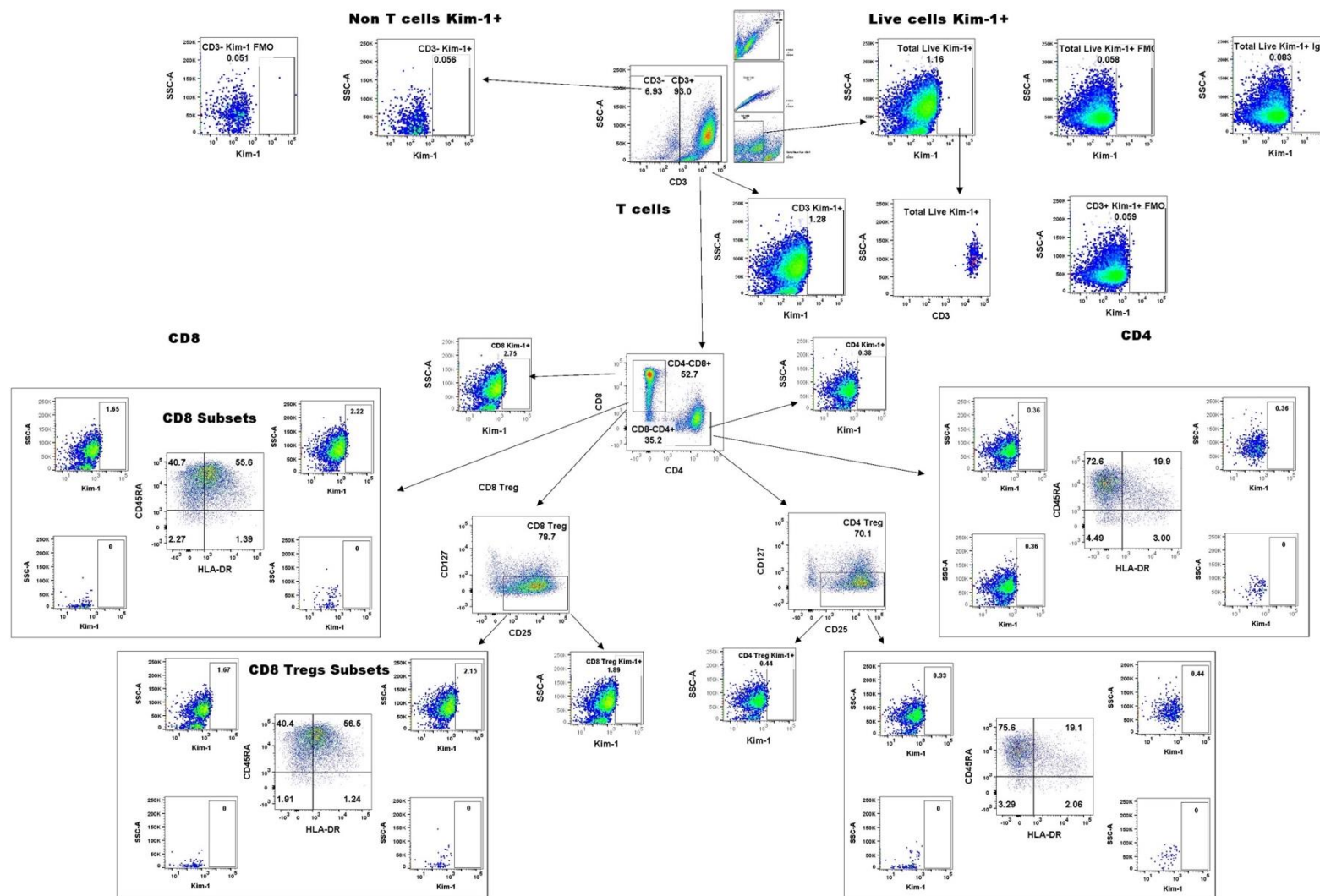
Suppl. Fig. 3. Glucose variability even in the absence of diabetes increases T cell Kim-1 expression in preclinical models. Male adolescent apolipoprotein E deficient mice (*ApoE*^{-/-}; 6 weeks of age) received four intraperitoneal (i.p.) injections of glucose (2g/kg; blue diamond symbols; N=10) or isovolumetric saline injections (saline; white diamonds; N=10), delivered two hours apart, to achieve plasma glucose variations which peaked between ~15-20mmol/L. **A-B** Kidney cortical gene expression by qPCR of (A) Collagen IV (*Col4a1*) or (B) Kim-1 (*Havcr1*). **C-D** Representative photomicrographs of kidney cortical Kim-1 (red) and nuclear (DAPI, blue) immunofluorescence in (C) saline injected or (D) diabetic mice. Scale Bar = 50 µm; x200 Magnification. **E-G** Flow cytometry analysis of live peripheral blood Kim-1 positive (Kim-1⁺) (E) T cells (Kim-1⁺CD3⁺); (F) CD4⁺ (Kim-1⁺CD3⁺CD4⁺CD8⁻) and CD8⁺ (Kim-1⁺CD3⁺CD8⁺CD4⁻) T cells as proportion of CD3⁺KIM-1⁺ T cells. Linear regression of (G) CD4⁺CD8⁻Kim-1⁺; (H) Characteristics of *ApoE*^{-/-} murine groups at the study end. Data are shown as mean SD or median (IQR) and tested using 1 way ANOVA/Tukey's post-hoc or Kruskal Wallis/Dunn's post-hoc testing. **P*<0.05; ***P*<0.01, ****P*<0.001 vs saline group, ###*P*<0.001 vs glucose group.



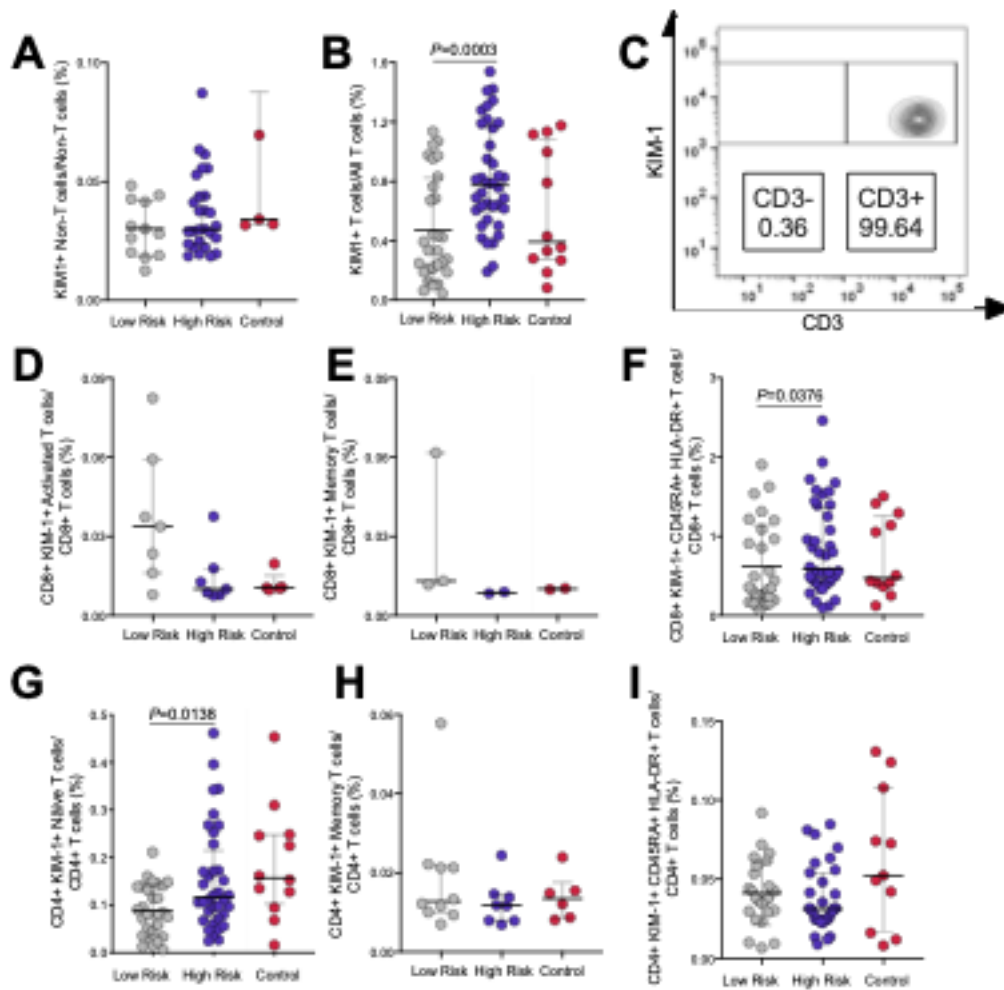
Suppl. Fig. S4. Gating strategy for KIM-1+ Peripheral Blood Mononuclear Cells in *ApoE*^{-/-} mice at risk for DKD.



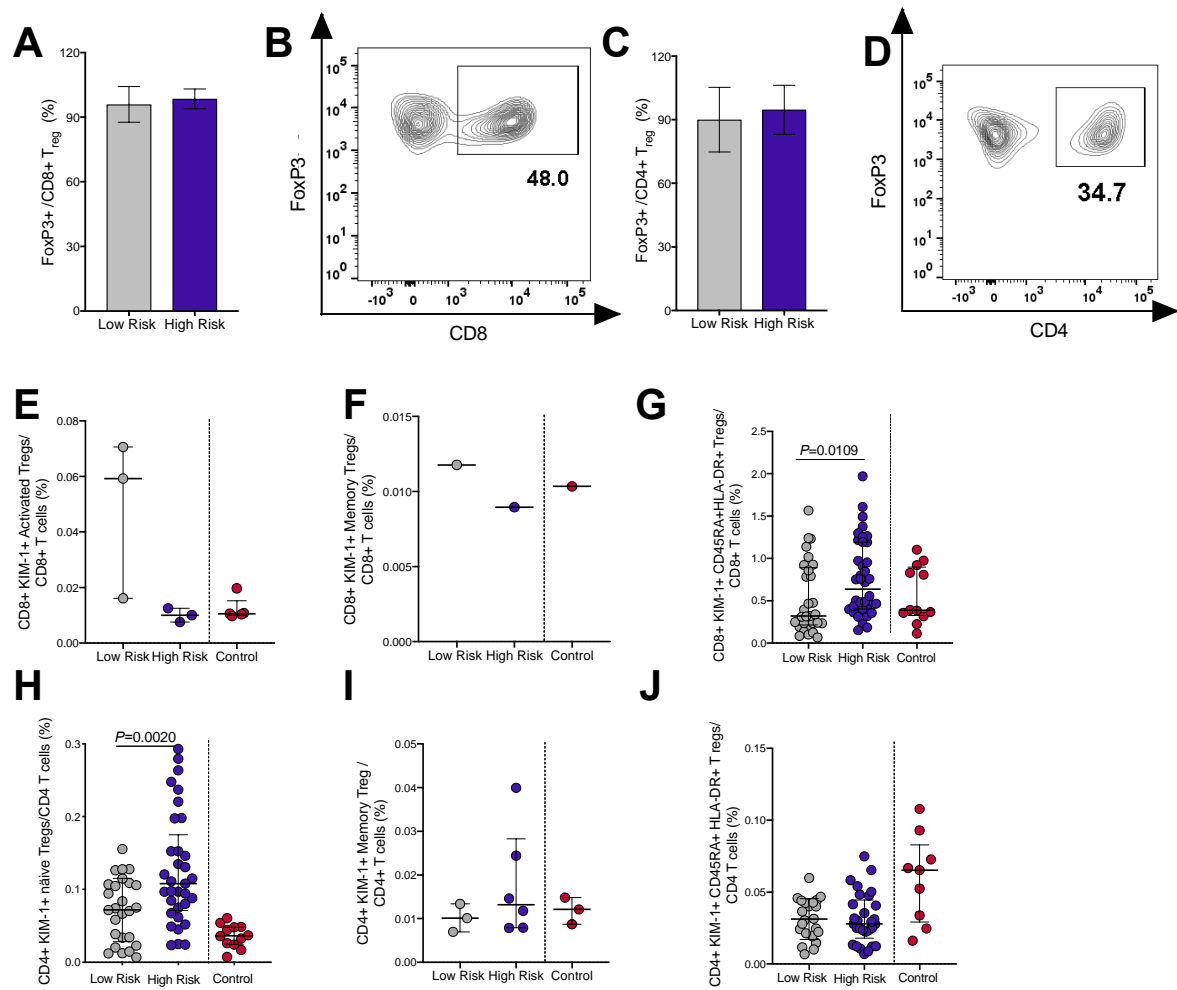
Suppl. Fig. S5. Peripheral blood mononuclear cell expression of *HAVCR1* (KIM-1 gene) in healthy human donors. Expression data was obtained from the gene expression omnibus (GEO) dataset GSE77671. Data were processed for background correction, normalization and log2-transformation within R and integrated into the Stemformatics platform [www.stemformatics.org] for visualization. $N=30$ donations from peripheral blood. The probe shown for *HAVCR1* is A_23_P347610.



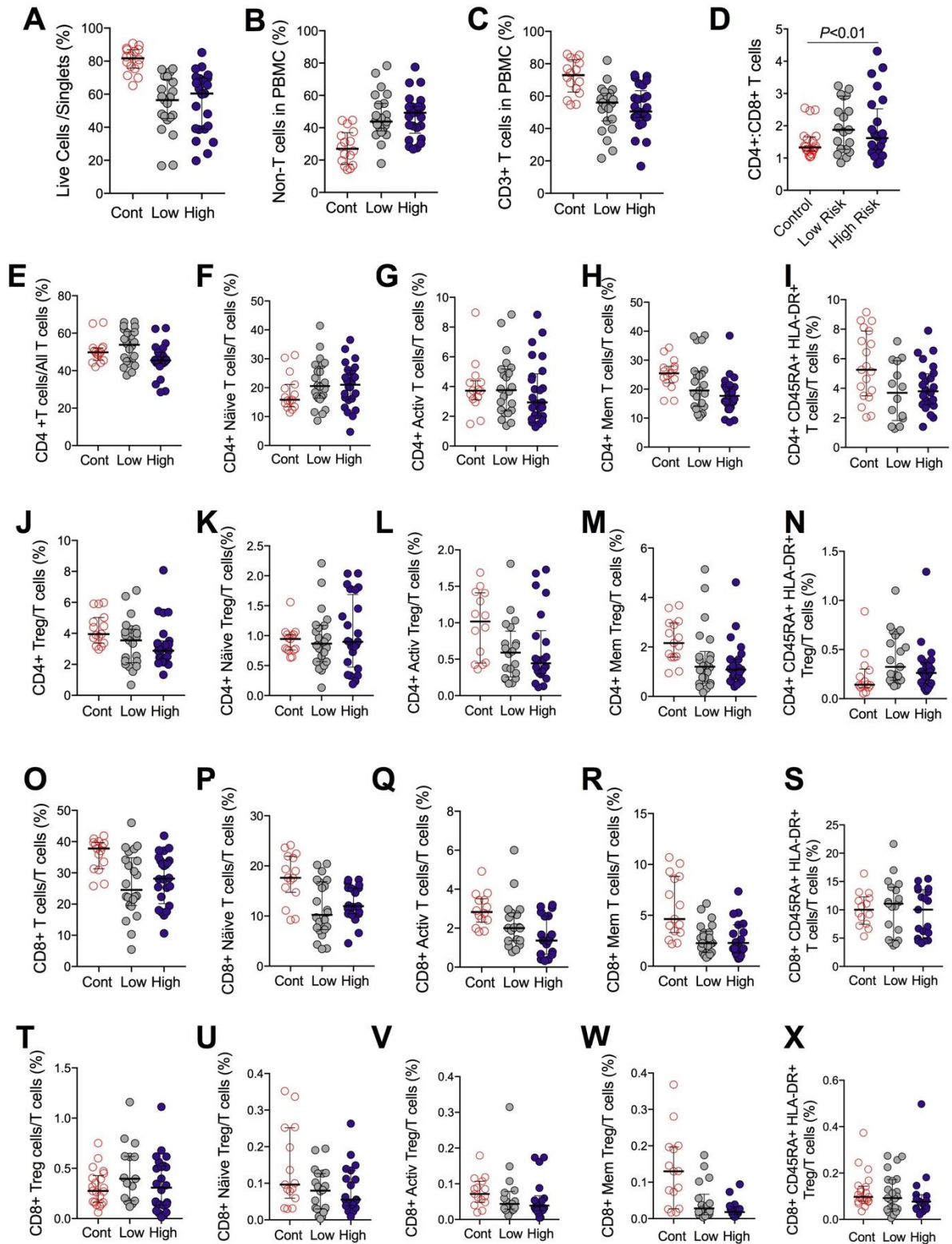
Suppl. Fig. S6. Gating strategy for Peripheral Blood Mononuclear Cells in youth at risk for DKD.



Suppl. Fig. S7. T conventional cell subset KIM-1 expression in youth at high risk for DKD. Youth (20.0 ± 2.8 yrs old) with type 1 diabetes were stratified by risk for DKD using tertiles of urinary albumin:creatinine ratio (uACR; Low Risk , $N=33$; Medium Risk, $N=33$; High Risk, $N=34$). KIM-1 expression on live peripheral blood mononuclear cells (PBMCs) is shown for low and high risk groups. Circulating PBMC proportions of live KIM-1+ (A) CD3⁻ KIM-1+ non-T cells and (B) CD3⁺KIM-1+ T cells. (C) Live KIM-1+ PBMC shown as a contour diagram. Live CD3⁺ T cells expressing CD4⁺ and CD8⁺ were sorted into specific subsets of (D) CD8⁺ activated T_{conv} (CD3⁺CD8⁺CD4⁺CD45RA⁻HLA-DR⁺); (E) CD8⁺ Memory T_{conv} (CD3⁺CD8⁺CD4⁺CD45RA⁺HLA-DR⁻); (F) CD3⁺CD8⁺CD4⁺CD45RA⁺HLA-DR⁺ T_{conv} double positive cells; (G) CD4⁺ naïve T_{conv} (KIM-1⁺CD3⁺CD4⁺CD8⁻CD45RA⁻HLA-DR⁻) T cells; (H) CD4⁺ Memory T_{conv} (CD3⁺CD4⁺CD8⁻CD45RA⁺HLA-DR⁻); or (I) CD3⁺CD4⁺CD8⁻CD45RA⁺HLA-DR⁺ T_{conv} double positive cells. KIM-1⁺CD4⁺ activated T_{conv} (KIM-1⁺CD3⁺CD4⁺CD8⁻CD45RA⁺HLA-DR⁺) T cells are not shown as they were present in negligible quantities; Data are shown as mean \pm SD or median (IQR) and tested using Student's T test or Mann Whitney U testing.



Suppl. Fig. S8. T regulatory cell subset KIM-1 expression in youth at high risk for DKD. Youth (20.0 ± 2.8 yrs old) with type 1 diabetes were stratified by risk for DKD using tertiles of urinary albumin:creatinine ratio (uACR; Low Risk, $N=33$; Medium Risk, $N=33$; High Risk, $N=34$). KIM-1 expression on live peripheral blood mononuclear cells (PBMCs) is shown for low and high risk groups. **A-D** FOXP3 expression in **(A)** CD8+ T regulatory cells (T_{reg}); **(B)** CD8+ T_{reg} as a Contour plot; **(C)** CD4+ T_{reg} and **(D)** as a Contour plot in CD4+ T_{reg} cells. Circulating live CD3+KIM-1+ T_{reg} expressing CD4+ and CD8+ were sorted into specific subsets of **(E)** CD8+ activated T_{reg} (CD3+CD8+CD4⁻CD25+CD127^{lo/-}CD45RA⁻HLA-DR⁺); **(F)** CD8+ Memory T_{reg} (CD3+CD8+CD4⁻CD25+CD127^{lo/-}CD45RA⁻HLA-DR⁻); **(G)** CD3+CD8+CD4⁻CD25+CD127^{lo/-}CD45RA+HLA-DR+ double positive T_{reg} cells; **(H)** CD4+ naïve T_{reg} (CD3+CD4+CD8⁻CD25+CD127^{lo/-}CD45RA⁻HLA-DR⁻); **(I)** CD4+ Memory T_{reg} (CD3+CD4+CD8⁻CD25+CD127^{lo/-}CD45RA⁻HLA-DR⁻); **(J)** CD3+CD4+CD8⁻CD25+CD127^{lo/-}CD45RA+HLA-DR+ double positive T_{reg} cells. KIM-1+CD4+ activated T_{reg} (CD3+CD4+CD8⁻CD25+CD127^{lo/-}CD45RA⁻HLA-DR⁺) are not shown as they were present in negligible numbers. Data are shown as mean \pm SD or median (IQR) and tested using Student's T test or Mann Whitney U testing.



Suppl. Fig. S9. White blood cell subsets in peripheral blood from youth with type 1 diabetes and varied risk for DKD. Peripheral leukocytes were obtained from youth (20.0 ± 2.8 yrs old) with type 1 diabetes with varying risk for DKD (uACR; Low Risk, $N=33$; Medium Risk, $N=33$; High Risk, $N=34$). (A) Live cells were quantified using flow cytometry into (B) Non T cell subsets (CD3⁻); (C) T cells (CD3⁺); (D) CD4+:CD8+ T cell ratio; (E) CD4+ conventional T cells (CD3+CD4+CD8⁺) and F-I the CD4+ conventional (T_{conv}) T cell subsets of (F) naïve T_{conv} (CD3+CD4+CD8⁻CD45RA+HLA-DR⁻); (G) activated T_{conv}

(CD3+CD4+CD8⁻CD45RA⁻HLA-DR⁺); **(H)** memory T_{conv} (CD3+CD4+CD8⁻CD45RA⁻HLA-DR⁻) and **(I)** CD3+CD4+CD8⁻CD45RA+HLA-DR+ T_{conv}. **(J)** CD4⁺ T regulatory (T_{reg}) cells (CD3+CD4+CD8⁻CD25+CD127^{lo/-}) sorted into **K-N** the CD4⁺ T_{reg} subsets of **(K)** naïve T_{reg} (CD3+CD4+CD8⁻CD25+CD127^{lo/-}CD45RA+HLA-DR⁻); **(L)** CD4⁺ activated T_{reg} (CD3+CD4+CD8⁻CD25+CD127^{lo/-}CD45RA-HLA-DR⁺); **(M)** CD4⁺ memory T_{reg} (CD3+CD4+CD8⁻CD25+CD127^{lo/-}CD45RA⁻HLA-DR⁻) and **(N)** CD3+CD4+CD8⁻CD25+CD127^{lo/-}CD45RA+HLA-DR+ T_{reg}. **(O)** CD8⁺ T_{conv} cells (CD3+CD8+CD4⁻) sorted into **P-S** the CD8⁺ T_{conv} subsets of **(P)** naïve T_{conv} (CD3+CD8+CD4⁻CD45RA+HLA-DR⁻); **(Q)** activated T_{conv} (CD3+CD8+CD4⁻CD45RA⁻HLA-DR⁺); **(R)** memory T_{conv} (CD3+CD8+CD4⁻CD45RA⁻HLA-DR⁻) and **(S)** CD3+CD8+CD4⁻CD45RA+HLA-DR+ T_{conv}. **(T)** CD8⁺ T regulatory (T_{reg}) cells (CD3+CD8+CD4⁻CD25+CD127^{lo/-}) sorted into **U-X** the CD8⁺ T_{reg} subsets of **(U)** naïve T_{reg} (CD3+CD8+CD4⁻CD25+CD127^{lo/-}CD45RA+HLA-DR⁻); **(V)** CD8⁺ activated T_{reg} (CD3+CD8+CD4⁻CD25+CD127^{lo/-}CD45RA-HLA-DR⁺); **(W)** CD8⁺ memory T_{reg} (CD3+CD8+CD4⁻CD25+CD127^{lo/-}CD45RA⁻HLA-DR⁻) and **(X)** CD3+CD8+CD4⁻CD25+CD127^{lo/-}CD45RA+HLA-DR+ T_{reg}. Data are shown as mean SD or median (IQR) and tested using 1 way ANOVA/Tukey's post-hoc or Kruskal Wallis/Dunn's post-hoc testing.