

Supplementary Files

Obesity-induced *miR-455* upregulation promotes adaptive pancreatic β -cell proliferation through the CPEB1/CDKN1B pathway

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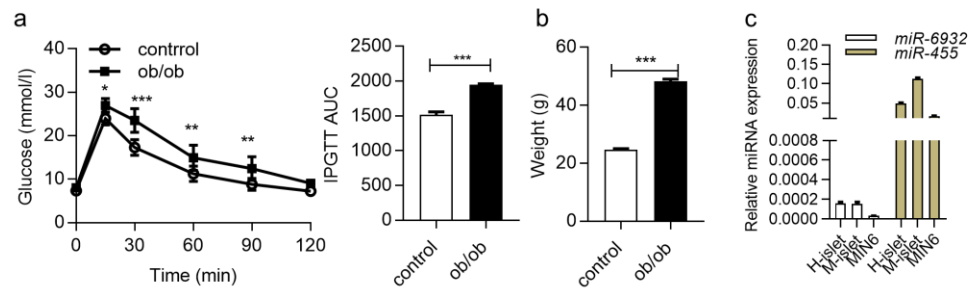
Table S2

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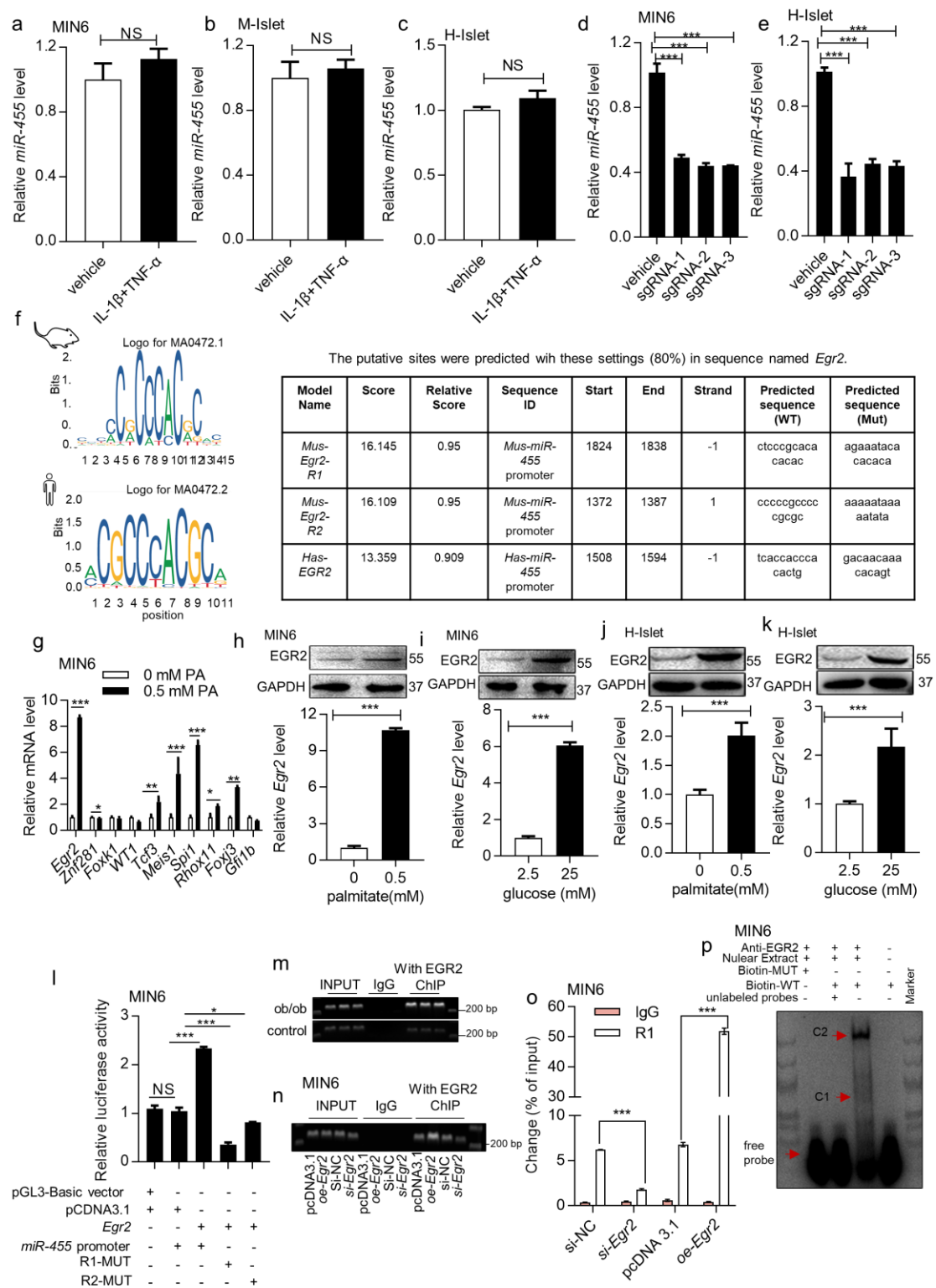
Table S5

Supplementary Figure 1



Supplementary Figure 1 (a) Blood glucose measurements during a glucose tolerance test in 8-week *ob/ob* mice and control mice ($n = 6$ mice per group). (b) The weight of 8-week-old *ob/ob* mice ($n = 6$ mice per group). (c) The expression levels of *miR-455* and *miR-6932* in human islets, mouse islets and MIN6 cells. The data are presented as the mean \pm SD. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

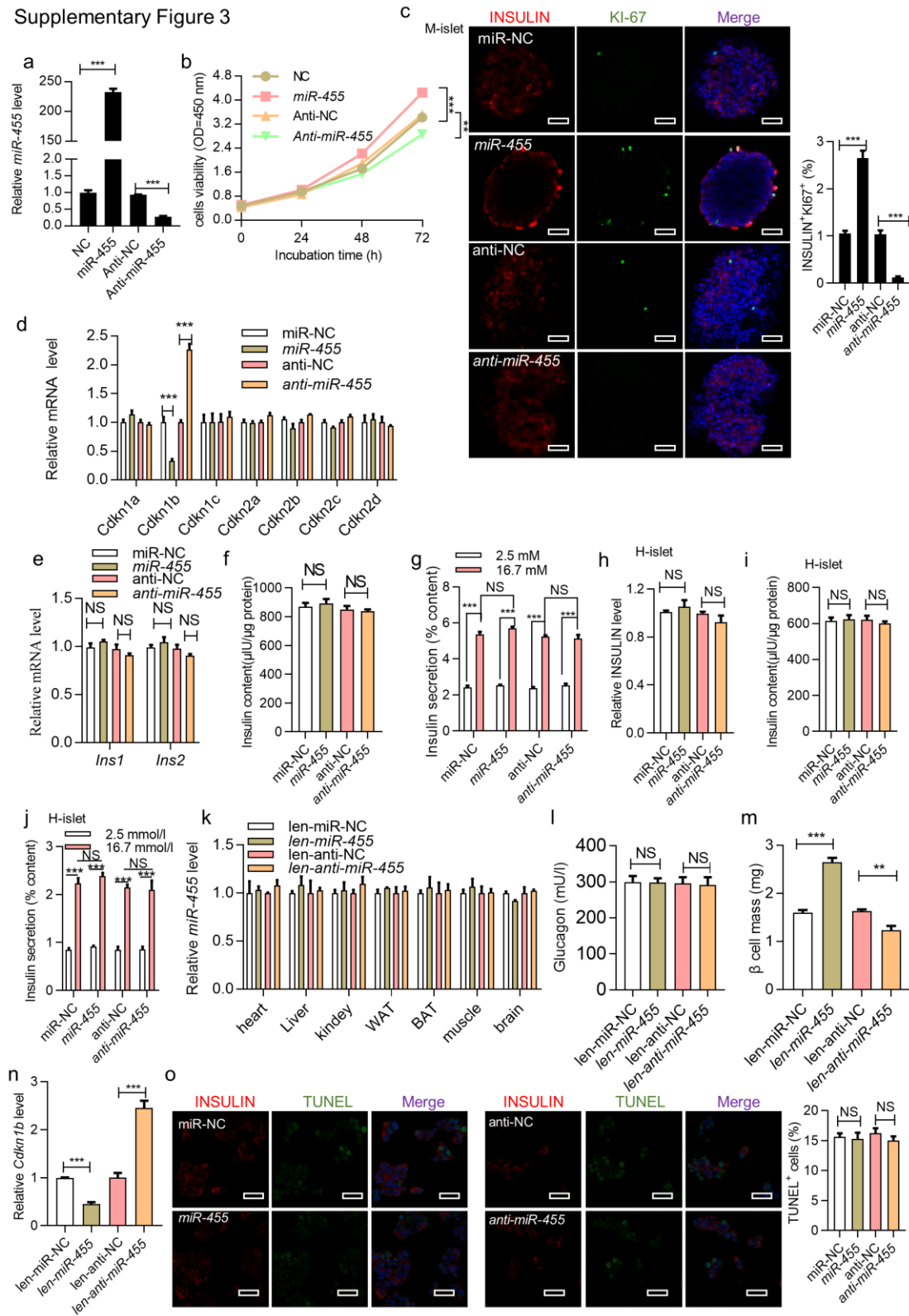
Supplementary Figure 2



Supplementary Figure 2 MIN6 cells (a), mouse primary islets (b, $n = 3$), and human islets (c) were incubated with a combination of interleukin-1 β (IL-1 β , 5 ng/ml) and tumor necrosis factor- α (TNF- α , 30 ng/ml) for 48 h, and qRT-PCR was performed to examine the *miR-455* expression level. (d-e) The expression level of *miR-455* in MIN6 cells (d) and in human islets (e) transfected

with *miR-455* sgRNA. (f) The binding sites of *Egr2* in the *mus-miR-455* and *human-miR-455* promoters predicted by JASPAR. (g) MIN6 cells were incubated with 0.5 mM palmitate for 48 h, and qRT-PCR was performed to examine the expression levels of transcription factors predicted by JASPAR. MIN6 cells and human islets were incubated with 0.5 mM palmitate or 25 mM glucose for 48 h, and the protein and mRNA levels of EGR2 were determined by western blotting and qRT-PCR (h-k), respectively. (l) Relative luciferase activity in MIN6 cells cotransfected with *Egr2* overexpression plasmid and a luciferase reporter containing either *miR-455 promoter* or *miR-455-MUT* (*Egr2*-binding sequence mutated). The data are presented as the relative ratio of Renilla luciferase activity to firefly luciferase activity. ChIP experiments were conducted to verify that *Egr2* binds to the promoter of *miR-455* in islets from obese mice (m). ChIP experiments were conducted to verify that *Egr2* binds to the promoter of *miR-455* in MIN6 cells transfected with *oe-Egr2* or *si-Egr2*, followed by RT-PCR (n) and qRT-PCR (o) assays. (p) Based on EMSA results, *Egr2* can directly bind to the *miR-455* promoter in MIN6 cells. C1 and C2 represent nuclear protein-*miR-455* probe complexes and nuclear protein-*miR-455* probe-anti-EGR2 complexes, respectively. The data are presented as the mean \pm SD. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

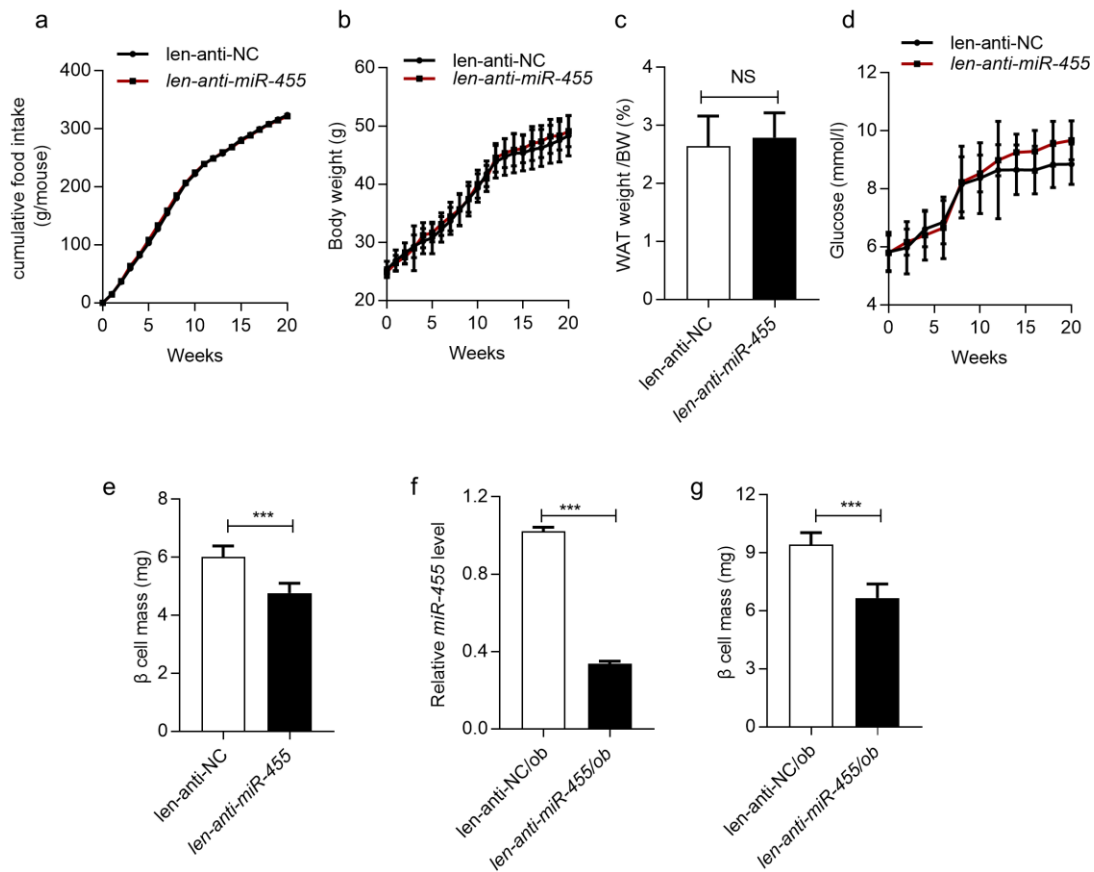
Supplementary Figure 3



Supplementary Figure 3 MIN6 cells were transfected with *miR-455* or *anti-miR-455* for 48 h, and qRT-PCR was performed to detect the *miR-455* expression level (a). (b) CCK-8 assays were carried out at 24, 48, and 72 h after transfection. (c) INSULIN⁺/KI-67⁺ cells were counted via immunofluorescence. Magnification: 20 x; scale bar, 20 μm. (d) The mRNA expression levels of

Cdkn1 and *Cdkn2* family genes. The *Ins1* and *Ins2* expression levels (e), insulin content (f) and insulin secretion (g) in MIN6 cells. INSULIN levels (h), insulin content (i) and insulin secretion (j) in human islets. (k) The *miR-455* expression levels in different tissues from *len-miR-455* or *len-anti-miR-455* mice ($n = 3$). (l) The glucagon level in serum from *len-miR-455* or *len-anti-miR-455* mice ($n = 9$ mice per group). (m) The β cell mass in *len-miR-455* or *len-anti-miR-455* mice ($n = 3$). (n) The mRNA levels of *Cdkn1b* in the islets of *len-miR-455* or *len-anti-miR-455* mice ($n = 3$). (o) INSULIN⁺/TUNEL⁺ cells were counted via immunofluorescence. Magnification: 20 x; scale bar, 20 μ m. The data are presented as the mean \pm SD. ** $p < 0.01$, *** $p < 0.001$.

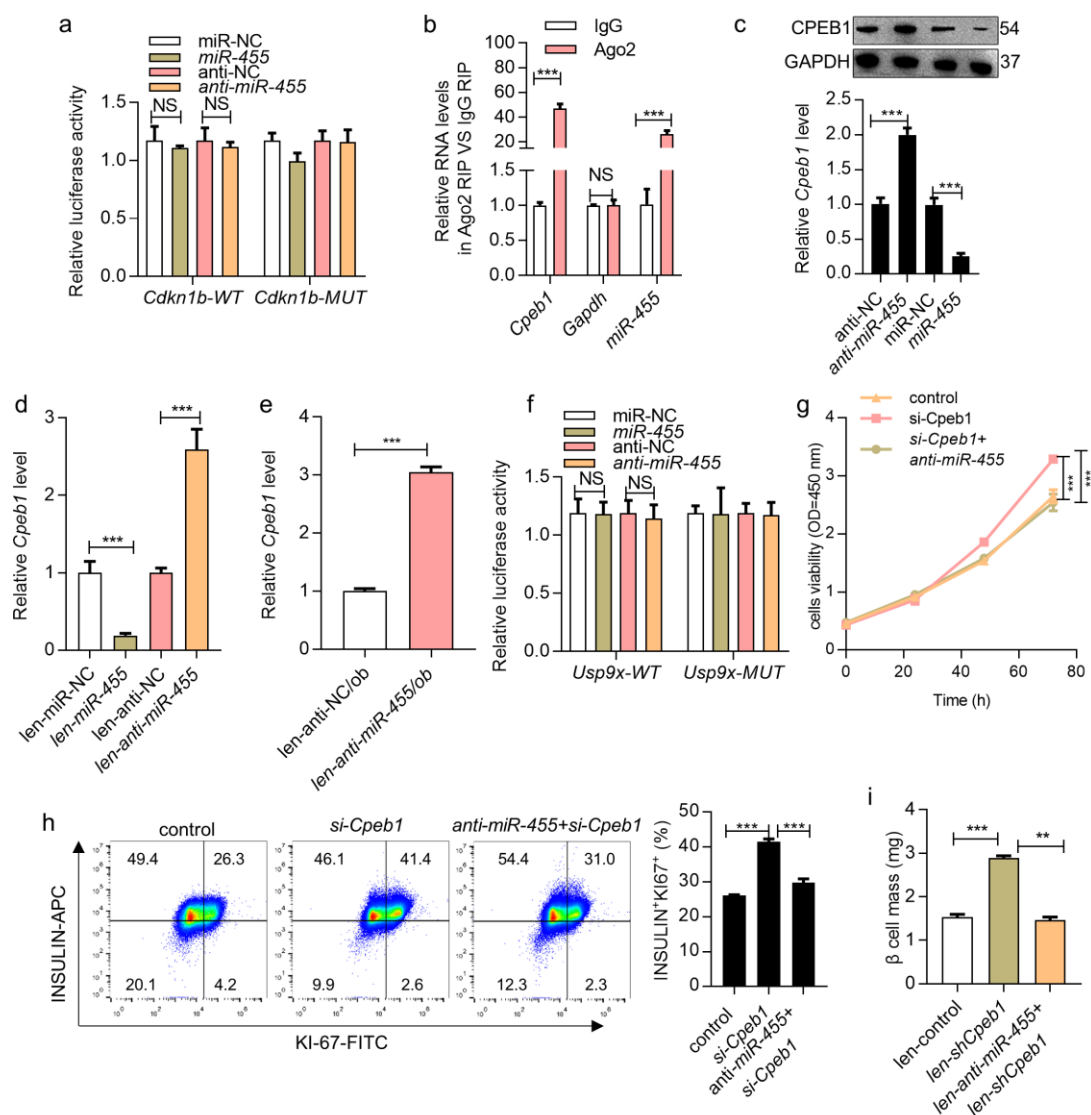
Supplementary Figure 4



Supplementary Figure 4 Eight-week-old male *len-anti-miR-455* mice and control mice (lentivirus-MIP (mouse insulin2 promoter)-LV3/H1 vector) were exposed to an HFD for 20 weeks.

Then, cumulative energy intake (a, $n = 9$ mice per group), changes in body weight (b, $n = 9$ mice per group), the white adipose tissue weight per body weight ratio (c, $n = 9$ mice per group) and fasting blood glucose level (FBG) (d, $n = 9$ mice per group) were measured. (e) The β cell mass of *len-anti-miR-455* mice was measured ($n = 3$). (f) *miR-455* expression levels in the islets of *len-anti-miR-455/ob* mice ($n = 3$). (g) The β cell mass in *len-anti-miR-455/ob* mice was measured ($n = 3$). The data are presented as the mean \pm SD. *** $p < 0.001$.

Supplementary Figure 5



Supplementary Figure 5 (a) Relative luciferase activity of MIN6 cells cotransfected with *miR-455* mimic and a luciferase reporter containing either *Cdkn1b-WT* or *Cdkn1b-MUT*. The data are presented as the relative ratio of Renilla luciferase activity to firefly luciferase activity. (b)

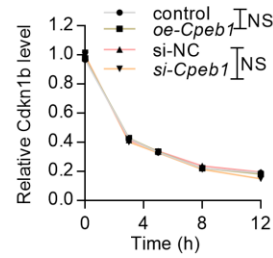
MIN6 cells were transfected with *Cpeb1* overexpression plasmid for 48 h. Then, MIN6 cell lysate was subjected to anti-Ago2 RNA immunoprecipitation (RIP), and the *miR-455* and *Cpeb1* expression levels were examined via qRT-PCR. *Gapdh* served as a control (nonspecific IgG served as a negative control). (c) CPEB1 mRNA and protein levels in MIN6 cells transfected with *miR-455* or *anti-miR-455*. (d-e) The *Cpeb1* mRNA level in islets from *len-miR-455* mice and *len-anti-miR-455/ob* mice ($n = 3$). (f) Relative luciferase activity of MIN6 cells cotransfected with *miR-455* mimic and a luciferase reporter containing either *Usp9x-WT* or *Usp9x-MUT*. The data are presented as the relative ratio of Renilla luciferase activity to firefly luciferase activity. (g) CCK-8 assays were carried out at 24, 48, and 72 h after transfection with *si-Cpeb1* or *si-Cpeb1* and *anti-miR-455*. (h) The INSULIN⁺/KI-67⁺ cells were counted via flow cytometry. (i) The β cell mass in *len-shCpeb1* mice was measured ($n = 3$). The data are presented as the mean \pm SD. ** $p < 0.01$, *** $p < 0.001$.

Supplementary Figure 6

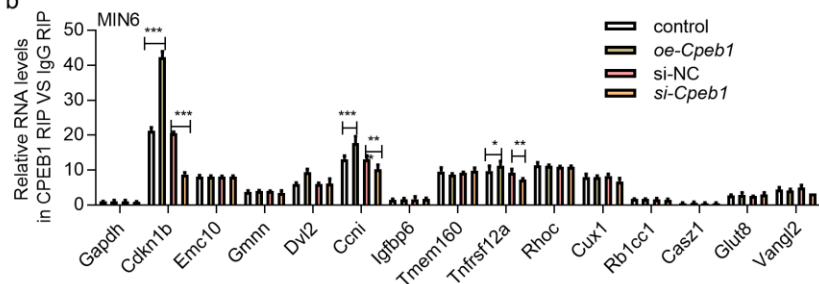
a

Gene name	ctaRAPID	RBPmap	Score
<i>Rb1cc1</i>	+	-	6.23
<i>Cdkn1b</i>	+	+	5.93
<i>Rhoc</i>	+	-	5.86
<i>Dvl2</i>	+	+	5.7
<i>Igfbp6</i>	+	-	5.59
<i>Gmn</i>	+	-	5.33
<i>Tmem160</i>	+	-	5.05
<i>Tnfrsf12a</i>	+	+	4.94
<i>Vangl2</i>	+	-	5.04
<i>Emc10</i>	+	-	4.89
<i>Ccni</i>	+	+	4.77
<i>Cas1</i>	+	-	4.66
<i>Cux1</i>	+	-	4.64
<i>GLUT8</i>	+	-	4.57

c

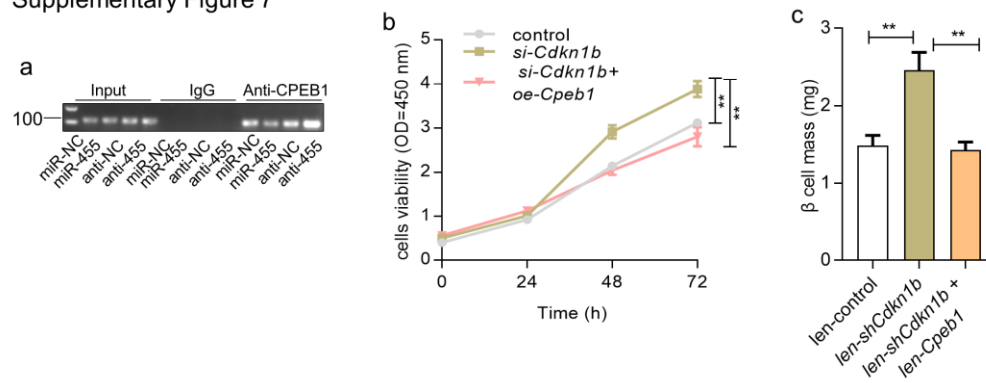


b



Supplementary Figure 6 (a) ctaRAPID and RBPmap were used to predict mRNAs that can bind to CPEB1. (b) MIN6 cells were transfected with *oe-Cpeb1* and *si-Cpeb1* for 48 h. Then, MIN6 cell lysate was subjected to anti-CPEB1 RNA immunoprecipitation (RIP), and the precipitated RNAs were examined via qRT-PCR. *Gapdh* served as a control (nonspecific IgG served as a negative control). (c) MIN6 cells transfected with *oe-Cpeb1* or *si-Cpeb1* were treated with actinomycin D (10 μ g/ml). RNA was isolated at various time points after the treatment and analyzed via RT-qPCR. The data are presented as the mean \pm SD. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

Supplementary Figure 7



Supplementary Figure 7 (a) RT-PCR-RIP was performed to verify the CPEB1–*Cdkn1b* interaction. (b) CCK-8 assays were carried out at 24, 48, and 72 h after transfection with *si-Cdkn1b* or *si-Cdkn1b* and *oe-Cpeb1*. (c) The β cell mass in *len-shCpeb1* mice was measured ($n = 3$). The data are presented as the mean \pm SD. ** $p < 0.01$.

Supplementary Spreadsheets:

Table S1 RNA isolated from islets of control mice and *ob/ob* mice, this table shows significantly changed miRNA (Log2 fold change ≥ 2). The mean expression of each group was presented in a log2 scale.

miR_name	control-read	<i>ob/ob</i> -read	log2(fold change)	p-value	FDR
mmu-miR-6932-3p	0.00	5.20	12.34	0.000899	0.018575
mmu-miR-455-5p	1075.12	15104.93	3.84	3.60E-08	2.13E-06
mmu-miR-677-3p	1.25	13.57	3.44	6.30E-05	0.002277
mmu-miR-6546-5p	1.29	12.19	3.25	0.000176	0.005328
mmu-miR-212-3p	815.84	7001.88	3.10	9.37E-20	2.44E-17
mmu-miR-132-3p	5648.74	41049.84	2.86	2.43E-21	7.91E-19
mmu-miR-132-5p	589.23	4146.32	2.81	3.84E-16	5.55E-14
mmu-miR-501-5p	91.11	636.67	2.80	2.31E-13	2.31E-11
mmu-miR-212-5p	516.13	3417.69	2.73	4.47E-15	5.81E-13
mmu-miR-1945	2.42	14.32	2.56	0.000368	0.008713
mmu-miR-6546-3p	2.31	11.77	2.35	0.002614	0.038215
mmu-miR-1983	38.48	185.64	2.27	2.38E-08	1.55E-06
mmu-miR-665-5p	5.71	25.86	2.18	9.37E-05	0.003047
mmu-miR-7093-3p	8.78	36.52	2.06	7.71E-05	0.002638
mmu-miR-690	62.28	256.98	2.04	1.63E-07	9.23E-06
mmu-miR-712-5p	3.44	14.07	2.03	0.002991	0.042759
mmu-miR-150-5p	3591.51	867.79	-2.05	1.25E-09	9.00E-08
mmu-miR-142a-3p	6707.08	1492.88	-2.17	4.24E-11	3.67E-09
mmu-miR-669o-5p	18.09	3.74	-2.28	0.000317	0.00792
mmu-miR-592-5p	101.53	18.73	-2.44	3.20E-08	1.98E-06
mmu-miR-216b-5p	5763.70	974.22	-2.56	2.46E-14	2.91E-12
mmu-miR-216a-5p	13489.70	2161.92	-2.64	2.81E-16	4.58E-14
mmu-miR-217-5p	6074.72	865.87	-2.81	1.54E-16	2.86E-14
mmu-miR-216b-3p	741.02	102.69	-2.85	5.42E-14	5.88E-12
mmu-miR-216a-3p	99.84	13.45	-2.89	4.48E-10	3.65E-08
mmu-miR-6969-5p	30.21	4.05	-2.90	1.22E-06	6.10E-05
mmu-miR-142a-5p	12379.31	1612.28	-2.94	3.99E-19	8.66E-17
mmu-miR-217-3p	297.30	38.14	-2.96	3.63E-13	3.37E-11
mmu-miR-150-3p	10.44	1.27	-3.03	0.000203	0.005812
mmu-miR-6969-3p	69.65	8.49	-3.04	2.76E-09	1.89E-07
mmu-miR-196b-5p	13.90	1.26	-3.46	1.28E-05	0.000522
mmu-miR-184-3p	4929.72	257.98	-4.26	1.16E-30	7.54E-28
mmu-miR-122-3p	191.54	3.09	-5.95	8.33E-28	3.61E-25
mmu-miR-122-5p	5805.91	84.06	-6.11	4.47E-52	5.81E-49

Table S2 The primers used in PCR (5'-3').

Gene name	Forward Primer	Reverse Primer
<i>oe-Cpeb1</i>	ctatagggagaccaagctgATGGCTTTCTCT CTGGAAGAAGC	gctgatcagcgggtttaaacTCAGTTCTTCTG GTTCTTCATTAGG
<i>oe-Egr2</i>	ctatagggagaccaagctgATGATGACCGCC AAGGCC	gctgatcagcgggtttaaacTCACGGTGTCCCT GGTTTCGAG
<i>Has-EGR2</i>	ctatagggagaccaagctgtATGATGACCGC CAAGGCC	gctgatcagcgggtttaaacTCAAGGTGTCC GGGTCCG
<i>oe-Cdkn1b</i>	ctatagggagaccaagctgATGTCAAACGTG AGAGTGTCTAACG	gctgatcagcgggtttaaacTTACGTCTGGCG TCGAAGGC
promoter- <i>miR-4</i> 55	gagctcttacgcgtgctagcATTGGCAGAGGA CAAGAACAGG	cagtaccggaatgccaagctGGCAGGCCCT ACTTCATG
promoter- <i>miR-4</i> 55-MUT	gatctttatgtgtgtgtTGTGGTATGAATCGCC TTGTAGA	acacacacataaagaTCCCGCAGGCACAC TCCA
<i>Has-miR-455</i> promoter	agaacatttctctatcgataGCTCGACTTGCCT GCCAGG	agcttacttagatcgcagatGATGACATAGGC CTTGAGGCA
MUS-EMSA-W T	CGGGAGTGTGTGTGCGGGAGTGTG	CACACTCCCGCACACACACTCCCG
MUS-EMSA-M UT	CGGGATGTGTGTGTATTCTTGTG	CACAAGAAATACACACACACCCCG
HAS-EMSA-W T	CTGTCCAGTGTGGGTGGTGACAATC	GATTGTCACCACCCACACTGGACA G
HAS-EMSA-M UT	CTGTCACTGTGTTTGTGTCCAATC	GATTGGACAACAAACACAGTGAC AG
<i>Cpeb1-WT</i>	TCGAGtgacatgtct gccaatTTg ttctgggtgg gcacattTgg taatcctgtc tGT	CTAGAcagacaggattaccaaatgtcccaccag aacaaaattggcagacatgtcaC
<i>Cpeb1-MUT</i>	TCGAGtgacatgtct gccaatTTCACtCgggtgTTACACCtttgg taatcctgtc tGT	CTAGAcagacaggattacaaaaGGTGTAaCa cccGgaGTGaaattggcagacatgtcaC
Cdkn1b-WT	TCGAGTgttactacctgtgtatatagttttatcttttactct gtagcacataaT	CTAGAttatgtgctacagagtaaaagataaaaactat atacacaggtagtacaaC
Cdkn1b-MUT	TCGAG TgttactacctgtgtatatagtGGCCGCcttttactctgt agcacataa T	CTAGAttatgtgctacagagtaaaagGCGGCC actatatacacaggtagtacaaC

Table S3 The sequence of siRNA or sgRNA.

SiRNA/sgRNA name	Forward promoter	
<i>si-Cpeb1</i>	GCAGCACACAGUCAGUAUUTT	AAUACUGACUGUGUGCUGCTT
<i>si-Cdkn1b</i>	CCCGGUCAAUCAUGAAGAATT	UUCUUCAUGAUUGACCGGGTT
<i>si-Egr2</i>	AAGGTATCATCAATATTGTGAGT	ACTCACAATATTGATGATACCTT
<i>miR-455-sgRNA-1</i>	CACCGGTGGCTCACTTCCCAAGC AG	AAACCTGCTTGGGAAGTGAGCC ACC
<i>miR-455-sgRNA-2</i>	CACCGGGTCGCCCGCGCCAGGC CCA	AAACTGGGCCTGGCGCGGGCGA CCC
<i>miR-455-sgRNA-3</i>	CACCGGCTGCATCCCTTGCTGCG TC	AAACGACGCAGCAAGGGATGCA GCC
<i>Has-si-EGR2</i>	AGCUGUCUGACAACAUCUACC	UAGAUGUUGUCAGACAGCUGG
<i>Has-miR-455 sgRNA-1</i>	CACCGACTAACAGCGGCTGCGA ACC	AAACGGTTCGCAGCCGCTGTTAG TC
<i>Has-miR-455 sgRNA-1</i>	CACCGAAGGGCAGTAACCTCGC CCG	AAACCGGGCGAGGTTACTGCCCT C
<i>Has-miR-455 sgRNA-1</i>	CACCGGCAGAAGGAAGCTCGCG GCT	AAACAGCCGCGAGCTTCCTTCTG CC

Table S4 The primers used in Real-time PCR (5'-3').

Gene	Forward Primer	Reverse Primer
<i>Mus-miR-455</i>	TAATCTGACTATGTGCCTTTGGACT	TATGGTTTTGACGACTGTGTGAT
<i>Has-miR-455</i>	TAATCTGACTATGTGCCTTTGGACT	TATGGTTTTGACGACTGTGTGAT
<i>Cpeb1</i>	AAGGATTGCTGGGACAACCAA	GGCCACGGGGAGATTCTTG
<i>Gapdh</i>	AGGTCGGTGTGAACGGATTTG	TGTAGACCATGTAGTTGAGGTCA
<i>Cdkn1b</i>	TCAAACGTGAGAGTGTCTAACG	CCGGGCCGAAGAGATTTCTG
<i>ChIP-R1</i>	AGTACAGTGCCTGGCTGTGTCTGCG	AACTCACGAGCACGGCTCAGCCCTG
<i>ChIP-R2</i>	GCTCCTCCCGGCGCCGTCG	TGGGTCCCCAAGTGTGACGCTG
<i>HAS-ChIP</i>	GGCCCTGGGTGACCCCTGAAGTATG	GCTGCGAACCCTGGAGTGCCAACATT
<i>Ins1</i>	CACTTCCTACCCCTGCTGG	ACCACAAAGATGCTGTTTGACA
<i>Ins2</i>	GCTTCTTCTACACACCCATGTC	AGCACTGATCTACAATGCCAC
<i>Zfp281</i>	CCTTCCCCCAGAGTATGGTTA	TCCTTCTTGAAAGTCATGTCCG
<i>Foxl1</i>	GAGCAGAGGGGTCACACTGAAC	CTTCCTGCGCCGATAAATTGC
<i>Tcf3</i>	CGCACCAGCAGTACAGATGAG	CAGCTTGGTCTGCGCCTTA
<i>Meis1</i>	GCAAAGTATGCCAGGGGAGTA	TCCTGTGTTAAGAACCGAGGG
<i>Spi1</i>	ATGTTACAGGCGTGCAAAATGG	TGATCGCTATGGCTTTCTCCA
<i>Rhox11</i>	TGAGCTTCTGTGAGGAAGAAATC	CCTCTTTCGGGATGCCTGTTA
<i>Foxj3</i>	AGCCTAACATCTATGGACTGGT	GGTCAAGGAGTGCATTCTTCTTA
<i>Gfi1b</i>	ATGCCACGGTCCCTTCTAGTG	GGAAGGCTCTGGTTCAGCAA
<i>Wt1</i>	GAGAGCCAGCCTACCATCC	GGGTCCCTCGTGTTTGAAGGAA
<i>Ecm10</i>	TCAATGGCCTCTACAGGGTCC	AGCTTCTGAACCATCAAGTGTC
<i>Igfbp6</i>	GCTGCTAATGCTGTTGTTTCGC	GCACTTAGGGCTGTAGACCC
<i>Tmem160</i>	CTCCGCAAGGCACATGAGA	AGACCCACCGCATAGGAGG
<i>Cas21</i>	CCTCGCCTTTCTTGGGCAA	CACTGGGCATCACAGAAGTCC
<i>Gmnn</i>	GGGAGCCCAAGAGAATGTGAA	CAAGCCTTTTGGCAACTCATTT
<i>Vangl2</i>	ACTCGGGCTATTCTTACAAGT	TGATTTATCTCCACGACTCCCAT
<i>Cux1</i>	GGGGGAACAGGTTTCCAATAC	AAGGACTGCATTAGCTGTTCGC
<i>Dvl2</i>	GGTGTAGGCGAGACGAAGG	GCTGCAAAACGCTCTTGAAATC
<i>GLUT8</i>	CCCTTCGTGACTGGCTTTG	TGGGTAGGCGATTTCCGAGAT
<i>Ccni</i>	CAGGACCTTTGGAAAACCAGAG	TTGGATGGGCTTTTACTGTAGC
<i>Rb1cc1</i>	GACACTGAGCTAACTGTGCAA	GCGCTGTAAGTACACACTCTTC
<i>Tnfrsf12a</i>	GTGTTGGGATTTCGGCTTGGT	GTCCATGCACTTGTCGAGGTC
<i>Rhoc</i>	ATGGCTGCGATCCGAAAGAAG	GCACGTAGACCTCTGGAAACT
<i>Cdkn1a</i>	CCTGGTGATGTCCGACCTG	CCATGAGCGCATCGCAATC
<i>Cdkn1c</i>	CGAGGAGCAGGACGAGAATC	GAAGAAGTCGTTTCGCATTGGC
<i>Cdkn2a</i>	CGCAGGTTCTTGGTCACTGT	TGTTACGAAAGCCAGAGCG
<i>Cdkn2b</i>	CCCTGCCACCCTTACCAGA	CAGATACCTCGCAATGTCACG
<i>Cdkn2c</i>	CCTTGGGGGAACGAGTTGG	AAATTGGGATTAGCACCTCTGAG
<i>Cdkn2d</i>	CTGAACCGCTTTGGCAAGAC	GCCCTCTCTTATCGCCAGAT
<i>Has-CPEB1</i>	GATGCAAATGACTTGTGCCTTG	GGCTGAGGAATCTGAGTCCTG

<i>Has-EGR2</i>	TCAACATTGACATGACTGGAGAG	AGTGAAGGTCTGGTTTCTAGGT
<i>Has-Cdkn1b</i>	AACGTGCGAGTGTCTAACGG	CCCTCTAGGGGTTTGTGATTCT
<i>β-Actin</i>	GGCTGTATTCCCCTCCATCG	CCAGTTGGTAACAATGCCATGT

Table S5 The antibodies used in Western blot, IHC and IF.

Protein	Catalog	Dilution for WB	Dilution for IHC and IF	FACS
INSULIN-647	Cell Signaling (c27c9)			1:50
KI-67 FITC	Biologend (652409)			1:50
EGR2	Abcam (ab108399)	1:1000		
CPEB1	Abcam (ab73287)	1:1000		
CDKN1B	Abcam (193379)	1:1000		
INSULIN	Servicebio (GB13121)		1:500	
GLUCAGON	CST (2760)		1:500	
Argonaute 2	CST (2897s)	1:1000		
KI-67	Abcam (ab1667)		1:500	
GAPDH	Abcam (ab181602)	1:1000		